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Comparison of Dynamics of Extracellular Accesses to the $\beta_1$ and $\beta_2$ Adrenoceptors Binding Sites Uncovers the Potential of Kinetic Basis of Antagonist Selectivity

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Abstract

From the molecular mechanism of antagonist unbinding in the $\beta_1$ and $\beta_2$ adrenoceptors investigated by steered molecular dynamics, we attempt to provide further possibilities of ligand subtype and subspecies selectivity. We have simulated unbinding of $\beta_1$-selective Esmolol and $\beta_2$-selective ICI-118551 from both receptors to the extracellular environment and found distinct molecular features of unbinding. By calculating work profiles, we show different preference in antagonist unbinding pathways between the receptors, in particular, perpendicular to the membrane pathway is favourable in the $\beta_1$ adrenoceptor, whereas the lateral pathway involving helices 5, 6 and 7 is preferable in the $\beta_2$ adrenoceptor. The estimated free energy change of unbinding based on the preferable pathway correlates with the experimental ligand selectivity. We then show that the non-conserved K347 (6.58) appears to facilitate in guiding Esmolol to the extracellular surface via hydrogen bonds in the $\beta_1$ adrenoceptor. In contrast, hydrophobic and aromatic interactions dominate in driving ICI-118551 through the easiest pathway in the $\beta_2$ adrenoceptor. We show how our study can stimulate design of selective antagonists and discuss other possible molecular reasons of ligand selectivity, involving sequential binding of agonists and glycosylation of the receptor extracellular surface.

Keywords

adrenergic receptors; drug design; G protein-coupled receptors; molecular dynamics; selectivity

A major challenge in drug design is to find a small molecule that selectively binds to its target receptor and does not cause unintended side-effects by binding to other similar receptors. When a high-resolution structure of a receptor is available, a structure-based drug design paradigm is applicable to identify not only a small molecule ligand with high binding affinity, but also with good selectivity. However, the binding site architecture of closely related receptor subtypes and subspecies are often highly homologous, making the search for highly selective drugs, which relies on docking of small molecules into the crystal structures of receptors, impractical. This is particularly evident in the design of selective orthosteric
agonists and antagonists in such a large and pharmaceutically important class of drug targets as the G protein-coupled receptors.

One strategy for improving selectivity is to account for differences in the ligand binding and unbinding pathways of closely related receptors caused by non-conserved residues outside the drug-binding site. In this work, we aim to explore the dynamic and kinetic causes of antagonist subtype selectivity in the \( \beta_1 \) and \( \beta_2 \) adrenergic receptors (AR). \( \beta_1 \)AR and \( \beta_2 \)AR represent one of the most extensively characterized subfamilies of the G protein-coupled receptors, which are expressed in many cell types and play a pivotal role in regulation of the cardiovascular, pulmonary, endocrine and central nervous systems (1). Antagonists of the adrenergic receptors (\( \beta \)-blockers) are hallmark drugs for treatment of ischaemic heart disease, hypertension and congestive heart failure (1,2). Although the primary cardiovascular use of \( \beta \)-blockers is antagonism of \( \beta_1 \)AR responses in the heart, their use may also result in antagonism of \( \beta_2 \)AR in airways, resulting in bronchospasm (1,2). To avoid this side-effect, \( \beta_1 \)AR-selective antagonists are required.

To identify the dynamic and kinetic bases of antagonist selectivity, we have studied the unbinding process of two selective antagonists, Esmolol, which is 76-fold selective to \( \beta_1 \)AR (3–5), and ICI-118551, which is 550-fold \( \beta_2 \)AR selective (6,7) (Figure 1), from human \( \beta_1 \)AR and \( \beta_2 \)AR, using a molecular dynamics approach. Given that a computer-aided drug design campaign requires fast evaluation of potential binders, and monitoring of ligand binding and unbinding requires a microsecond time scale that is not affordable in a high-throughput level, we accelerated unbinding by applying an external force to pull the antagonist from the binding site in several directions using steered molecular dynamics (sMD). Through the use of multiple sMD simulations of ligand unbinding events, the statistical importance of unbinding pathways, as well as specific residue interactions important to them, can be analysed and key receptor conformations possessing characteristic interactions can be exploited in future drug design efforts.

Recent simulations of the unbinding pathways of the non-selective inverse agonist Carazolol from \( \beta_2 \)AR using the random acceleration molecular dynamics method have shown that unbinding occurs primarily through the extracellular region of \( \beta_2 \)AR and only rarely through transmembrane helices, suggesting that pathways through the extracellular surface provide a specific route to ligand entry (8). To further investigate this phenomenon, we have performed simulations in which the selective antagonists are pulled from the binding site to the extracellular surface along three directions: one perpendicular to the membrane and two lateral paths. We use the estimated free energy change of ligand unbinding based on the preferable pathway to correlate the experimentally observed ligand selectivity. From monitoring the impact of different interactions on the process of ligand unbinding, we delineate common and uncommon features of the unbinding process between two receptors and two ligands, along with their implications for structure-based drugs design studies. We also compare our results with the recent publication of non-selective ligand binding to \( \beta_2 \)AR using conventional MD (9) and non-selective ligand unbinding from \( \beta_1 \)AR and \( \beta_2 \)AR using steered MD (10). Finally, we discuss other potential reasons for different ligand binding and unbinding pathways between the receptors, which can account for structural constraints in the design of selective small molecule modulators.

**Methods**

**Preparation of ligand-protein complexes for simulations in the hydrated phospholipid bilayer**

To be consistent with ligand selectivity analysis in the human adrenergic receptors, we chose the available high-resolution crystal structure of the human \( \beta_2 \)AR (2.4 \( \text{Å} \)) (PDB ID: 78x747).
2RH1) and the 3D model of human \( \beta_1 \)AR was constructed based on this structure using the PRIME MODULE 2.2\(^a\) of the SCHRODINGER SUITE 9.0\(^b\). The 3D models of receptors were prepared for ligand docking studies using the Schrodinger protein preparation utility. The docking of ICI-118551 and Esmolol into the structures of \( \beta_1 \)AR and \( \beta_2 \)AR was performed with Glide 5.6\(^c\) with the Extra Precision algorithm. Ligands were processed using LoPro 2.4 with the OPLS 2005 force field\(^b\) (11,12). The investigated antagonists were close structural analogues of the ligands in the crystal structures, thus the docking poses similar to the crystal structures were quickly identified in the docking studies. Concurrent to this study, the crystal structure of \( \beta_2 \)AR in complex with ICI-118551 was published, which exhibit a nearly identical binding pose of ICI-118551 to our docking pose (13). The superposition of the crystallographic and computational coordinates of the binding site with ICI-118551 is shown in Figure S1 of the Supporting Information. The System Builder of Maestro 9.0 was used to embed four ligand-receptor complexes and the unoccupied receptors into a bilayer composed of 1-palmitoyl-2-oleoylphosphatidylcholine, solvate the extracellular and intracellular sides of the bilayer, neutralize the biomolecular system and preserve physiological pH using 0.15 M sodium chloride concentration. The final biomolecular systems contained around 75 000 atoms.

**Biomolecular dynamics**

Conventional and sMD were performed using the DYNAMOND package 2.2 (14). The temperature and pressure were controlled at 300 K and 1 atm, respectively, using the Berendsen algorithm. Long-range electrostatics was calculated by means of the Particle Mesh Ewald Method (15). The OPLS2005 force field was used for all simulations\(^b\) (11,12). The biomolecular systems were equilibrated using the following steps: (i) energy minimization (2000 cycles) of the solvent and lipid bilayer, while the ligand-receptor complex was frozen, using the conjugate gradient algorithm up to a convergence threshold of 0.5 kcal/mol/Å; (ii) heating of the system to 10 K in the NVT ensemble over 12 psecconds; (iii) heating of the system from 10 to 300 K in the NPT ensemble over 12 psecconds; (iv) equilibration for 7–9 nsecconds at 300 K in which the complexes were still frozen; (v) equilibration for 2 nsecconds in which the protein backbone was restrained; (vi) equilibration for 2 nsecconds with no restraints. Following equilibration, the 30 nsecconds production runs were carried out in the NPT ensemble. The steered MD simulations were tested with a pulling speed of 0.005, 0.01, 0.015, 0.02 and 0.03 Å/pseccond and a spring constant of 4.5, 4, 3.5, 2.5 and 1.5 kcal/mol/Å\(^2\) (16,17). The pulling speed of 0.02 Å/pseccond and spring constant of 4.0 kcal/mol/Å\(^2\) were selected for the detailed studies presented below. Ten pulling simulations in each chosen direction were performed with different initial velocities. Figure 2 was prepared in PyMOL 0.99\(^d\) and movies were made in Maestro\(^b\).

**Analysis of MD trajectories**

Hydrogen bond occupancy along simulations trajectories was computed in VMD 1.8.9 (18). Hydrogen bonds with a defined threshold for distance of 3 Å and angle of 35° were calculated in VMD 1.8.9 (18). Carbon atoms at distance 1.4–5 Å were selected for hydrophobic contacts in Maestro of the SCHRODINGER SUITE 9.0\(^b\). Water-mediated contacts at a distance of 3 Å with a minimum donor angle of 120° and a minimum acceptor angle of 90° were computed in Maestro. The radius of gyration of the lateral channel involving helices 5, 6 and 7 was calculated by selecting Ca atoms of F193, T195, H296 and N293 with VMD

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\(^a\)PRIME 2.0. (2008) New York, NY, USA: Schrödinger, LLC.

\(^b\)GLIDE 5.6. (2009) New York, NY, USA: Schrödinger, LLC.

\(^c\)LoPro 2.4. (2009) New York, NY, USA: Schrödinger, LLC.

\(^d\)The PyMOL Molecular Graphics System 0.99. (2002) San Carlos, CA, USA: DeLano Scientific, LLC.
(18). The work required pulling antagonists from the adrenergic receptors and ΔPMF were calculated using the VMD scripts.

Grid generation

The hydrogen bond acceptor and hydrophobic probes for the selected snapshots from sMD trajectories were calculated in S


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AP V2.5 of Schrodinger®. The probes were placed in the receptors at a radius of 6 Å from the ligand in the selected snapshot of the MD trajectory.

Results and Discussion

Dynamics of the extracellular side of the β1 and β2 adrenergic receptors with the selective ligands

We started our study with the performance of conventional molecular dynamics (cMD) simulations of the human β1 and β2 adrenergic receptors (AR) with docked Esmolol and ICI-118551 to initiate comparison of dynamics of the receptor extracellular cavity and ligand binding modes in each receptor, to define the ligand pulling directions from the receptor and to produce the starting coordinates for ligand unbinding simulations. The protein conformations stabilized following 2 nseconds of equilibration, at which point the root-mean-square deviations of protein backbone and ligand heavy atoms converged around 2 and 0.9 Å for the Esmolol-β1AR complex, 2.5 and 0.6 Å for the ICI-118551-β1AR complex, 2.1 and 0.6 Å for the Esmolol-β2AR complex and 1.9 and 0.7 Å for the ICI-118551-β2AR. The 30 nseconds cMD simulations of the complexes revealed higher fluctuation of the extracellular cavity, involving the extracellular regions of the helices and loops, in β1AR than in β2AR. As expected, the complex of the receptors with the selective antagonist was more rigid than the complex with the non-selective antagonist. The root-mean-square fluctuations of the Cα atoms of the extracellular cavity for two receptors are shown in Figure S2 in the Supporting Information. In Figure 2, we show one equilibrated snapshot of the extracellular cavity for each receptor, displaying amino acid residues of the extracellular side, together with the schematic representation of the antagonist binding site depicting the key residues involved in the interactions with the ligands.

In β1AR, a salt bridge between E205EL2 and R351EL3 was observed, with occupancy of more than 20% in each simulation, that dynamically connects the second extracellular loop (EL2) and the third extracellular loop (EL3) from the lateral side of the extracellular cavity in β1AR (Figure 2A). This salt bridge has also been observed in recent simulations of human β1AR by Gonzalez and co-workers (10) using the CHARMM force field. It is likely unique to the human β1AR as the residue corresponding to E205EL2 in turkey β1AR (Q188) is located 3.5–10 Å from the arginine side chain in the available crystal structures (19–21). In β2AR, these residues are substituted to E180EL2 and D300EL3.

Residues D192EL2 and K3057.46 (The Ballesteros and Weinstein nomenclature in superscript (22)), in β2AR form a salt bridge that splits the β2AR extracellular cavity into two sub-cavities (Figure 2B). This salt bridge, which is present in five of seven crystal structures of inactive β2AR bound to ligands (13,23,24), has an occupancy near 80% in simulations of the receptor complexes with Esmolol and ICI-118551. This is twice more than in our simulations of the ligand-free receptor, suggesting that antagonists stabilize this interaction, which likely holds the receptor in the inactive state. Although site-directed mutagenesis of D192EL2 and K3057.46 with emonstration of the role of these residues in ligand binding has not been documented in the literature, the different influence of ligands on the stability of the D192EL2-K3057.46 salt bridge has been recently described by an NMR
study via monitoring different resonances of radiolabeled K305 in the presence of agonists and antagonists (25). This mechanism of receptor modulation by extracellular salt bridges through their breakage and appearance was observed in the free fatty acid receptor 1 using mutagenesis and molecular modelling (26). β1AR misses this salt bridge as K305 is substituted to D356.

Hydrogen bonds between the secondary amine and β-hydroxyl group of both ligands with D138/113, N363/312 and Y367/316 (Figure 2C,D) were preserved during simulations with β1AR and β2AR, while the aromatic ring of ICI-118551 was buried deeper in the pocket than Esmolol in both receptors. The ester group of Esmolol was engaged in hydrogen bonding with N344/293 and S229/204 in 40% and 8% of the simulations time in β1AR, while in β2AR this was reduced to 13% and 3% of the simulation time. The varying interactions of N344/293 are owing to different accessibilities of N344/293 in the receptors; N344/293 is exposed to the extracellular cavity with χ mainly in the g-rotameric state, (171 ± 8°) in β1AR, whereas it is buried into the receptor with χ in the t-rotameric state, (−87 ± 25°) in β2AR. These different states are owing to a hydrogen bond between N293 and Y308 in β2AR, shown in the available crystal structures (13,21,22,26), that is disrupted by the substitution of Y308 to phenylalanine in β1AR (Figure 2C,D). Our simulations show that this hydrogen bond is formed more than 50% of the time in the complexes of β2AR with Esmolol and ICI-118551, reinforcing the non-direct role of the N293-Y308 interaction in ligand selectivity. The impact of N293-Q/L/A mutations in the reduction of β2AR ligand selectivity and, interestingly, the increase ligand selectivity for N293F has been shown by mutagenesis studies (29–31). This hydrogen bond was constantly monitored in the MD simulations of β2AR in the complex with carazolol by Vanni and co-workers (32). In the unoccupied receptor, the N293-Q/L/A hydrogen bond is present in only 20% of the simulations time, suggesting that ligand binding facilitates this interaction.

The final snapshots of the 30 nsec sMD simulations were used as starting points for sMD calculations. To investigate the impact of structural and dynamics differences of the extracellular cavity in ligand recognition, we pulled antagonists along three distinct directions, one perpendicular to the membrane (path B) and two lateral – one to the side of helices 1, 2 and 7 (path A) and the other to the side of helices 5, 6 and 7 (path C) (Figure 2).

**Pulling of selective antagonists from the β1 and β2 adrenergic receptors**

We used a steered MD algorithm to pull the antagonists from the binding pocket over 2 nsec s of simulation time. Pulling was induced along a particular pathway by the introduction of an external biasing potential along a predefined vector (as shown in Figure 2). The choice of the receptor and the antagonist pulling atoms (Figure 1) determined three ligand unbinding pathways, where the pulling directions were defined by the centre of mass of the receptor atoms are shown in Table 1. We tested simulations at varying pulling velocities and spring constants, which all exhibited unbinding in a similar fashion, and chose sMD parameters to balance computational cost with simulation accuracy (see Methods section). To investigate the pulling mechanics of selective antagonists, Esmolol and ICI-118551 from β1AR and β2AR in chosen directions, we calculated the rupture force profile along the pathways. Figure 3 shows the average force profile for the perpendicular and two lateral pathways of the receptors releasing either Esmolol or ICI-118551 projected onto the unbinding simulation time and the ligand separation distance from the original position. For each pathway, the average values were taken from ten trajectories simulated with a pulling velocity of 0.02 Å/ps and a spring constant of 4.0 kcal/mol Å^2.

Generally, complete unbinding of the ligand occurred within 1–1.5 nsec s. Although the receptors have a similar pattern of the force profile graphs between the pathways, there are notable differences in the size and dynamics of the force peaks, suggesting distinct
properties of the unbinding process. In β₁AR, the average force peak in path B had lower values than in paths A and C for both Esmolol and ICI-118551, while in β₂AR the force peaks along path C are lower than in paths A and B. To estimate the work performed by the antagonist during unbinding along chosen pathways we integrated the force curve over the ligand separation distance from the ligand original position (Figure 3). The computed work highlights the different preference in unbinding pathways and pulling of selective and non-selective antagonists between the receptors. In particular, less work is required to pull antagonists from path B in β₁AR and, among antagonists, more work is required to pull the β₁AR-selective antagonist in chosen pathways; less work is needed to pull the antagonists from path C and similarly, the β₂AR-selective antagonist is required more work to apply in β₂AR. Jarzynski’s equality states that the average work applied on the replicated ensembles can represent, in the first approximation, the overall change in the free energy between bound (a) and unbound (b) state or the potential of mean force (ΔPMF) along a reaction coordinate:

$$\exp(-\beta\Delta PMF) = \langle \exp(-\beta W_{a\rightarrow b}) \rangle_{ave}$$

where $\beta$ is $1/(k_bT)$, $k_b$ is Boltzmann constant and T is the temperature in Kelvin. Because in path B of β₁AR and path C of β₂AR, the ligands are needed the smallest work to perform, we use these pathways to compare ΔPMF with the experimental ligand selectivity. Although the calculation of the free energy change accurately requires numerous replicates (100–1000), we nevertheless hope that our multiple but still few simulations will provide a relative estimate of the free energy of ligand unbinding and binding. We repeated unbinding simulations for the preferable pathways (path B in β₁AR and path C in β₂AR) another ten times and calculated ΔPMF using the Jarzynski’s equality formula. The ΔPMF of Esmolol and ICI-118551 were −7 and −6 kcal/mol in path B of β₁AR and −6.3 and −7.9 kcal/mol in path C of β₂AR, respectively. The ΔPMF of ligands confirms antagonist preferences between the receptors. The calculated ΔPMF values from the available experimental constants of inhibition ($K_i$) (Figure 1) for Esmolol, were −7.3 and −6.8 kcal/mol and, for ICI-118551 were −10 and −12.6 kcal/mol, in β₁AR and β₂AR, respectively. The ΔPMF of unbinding from β₂AR is slightly lower from experimental calculated values, suggesting the importance of additional replicates to increase accuracy. Below, we compare unbinding of each selective antagonist from both receptors with characterization of the specific interactions that contribute to ΔPMF.

**Pulling from the β₁ adrenergic receptor**

Because path B required the smallest work to apply, we hypothesized that specific molecular features along this path might result in the experimentally observed ligand specificity of selective and non-selective antagonists. To examine this, we compared the unbinding of antagonists along path B and decomposed the rupture force into specific interactions, involving hydrogen bonds, hydrophobic, and water-mediated contacts, the results of which are shown in Figure 4 for one characteristic trajectory. From pharmacological studies, Esmolol binds in β₁AR more tightly than ICI-118551, thus more work is required to pull it out from the receptor, as indicated in the work profiles. After some jiggling resulting from the bias potential, involving the iterative breakage and appearance of the hydrogen bond between the ester group and N344 6.55, Esmolol ultimately lost its first hydrogen bond with the receptor via water-bridged interactions at early as 350 pseconds, corresponding to the first peak in the force profile. After some jiggling resulting from the bias potential, involving the iterative breakage and appearance of the hydrogen bond between the ester group and N344 6.55, Esmolol ultimately lost its first hydrogen bond with the receptor via water-bridged interactions at early as 350 pseconds, corresponding to the first peak in the force profile. This facilitated the movement of the aromatic ring of Esmolol towards the extra-cellular surface. Around 650 pseconds, the secondary amine and β-hydroxyl group of the ligand broke hydrogen bonds with N363 7.39 and Y367 7.43, and the ligand approached the extracellular surface while still retaining the salt bridge with D138 3.32.
in the binding site. The centre peak in the force profiles corresponds to the breakage of the salt bridge at the binding site, which happens after a minimum of 800 pseconds. After breakage of the interaction with D138^{3.32}, the amine of the ligand established interactions with D217^{EL2} in multiple trajectories and occasionally with D356^{EL3}, followed by diffusion into the water within 250 pseconds. In the ligand binding process, these two negatively charged residues likely serve as beacons that recognize the charged amino group of a ligand within the solvent environment. In the case of ICI-118551, the applied force caused the aromatic moiety to move towards the receptor surface, which was followed by the breakage of interactions with the original binding site residues. The unbinding process occurred quicker for ICI-118551 than for Esmolol.

The disruption of ligand-receptor interactions followed a similar pattern in other pulling directions. The ester group, released from its original interactions with binding site residues in β_{1}AR, formed hydrogen bonds with T220^{EL2} at 400–550 pseconds, followed by forming interactions with K347^{6.58} at 600–700 pseconds that lasted for 200 pseconds in seven of the simulation trajectories. After breakage of water-bridged interactions with K347^{6.58}, the ester group formed its last interaction with the backbone of F218^{EL2}. This profile of continuous interchange of hydrogen bond interactions between the ester and the receptor along the unbinding pathway is shown in Figure 5 and Movie S1 (where the role of K347^{6.58} is highlighted). We also observed direct and water-bridged interactions between the ester group and K347^{6.58} in five trajectories along path A. In contrast, ICI-118551 did not form any hydrogen bonds with K347^{6.58}. In both cMD and sMD simulations, K347^{6.58} moved freely in the cavity for a majority of the time and formed a salt bridge with E352^{EL3} in only 8% of the simulation time. Therefore, we propose that K347^{6.58} might act as a ‘hydrogen bond translator’ that interacts with a specific group of the ligand and facilitates the movement of the selective antagonist between the receptor extracellular surface and the binding site, along with preventing the exit of the molecule from the binding cavity. Potentially, during the ligand binding process, this residue might also facilitate Esmolol to enter the binding cavity with an orientation that would result in having optimal interactions with the binding site residues.

Antagonists formed interactions with aromatic and aliphatic residues for 53% and 47% of the simulation time, correspondently. Helix 2 has been proposed to play a role in β_{1}-selectivity of (−)-RO363 (33), and it is observed that the non-conserved I118^{2.64} held interactions with the ligand in 10% of the unbinding time, suggesting the potential role of I118^{2.64} in ligand selectivity, which was not found for (−)-RO363. During unbinding, the side chains of F218^{EL2} and F359^{7.35}, which form the gate to the extracellular cavity, moved away from each other, increasing the centre of mass distance between the aromatic rings from 8 to 12 Å (Figure 6A,B). Once the ligand exited the receptor, their separation distance decreased to a minimum of 5 Å, similar to the value observed in the unoccupied receptor (Figure 6C). This suggests that these two phenylalanines form gates that open and close during ligand binding, discriminate whether the ligand can enter into the binding cavity, and if so, guide the ligand into the receptor.

As the antagonists were pulled from the binding cavity, water-mediated interactions formed to replace protein-ligand interactions (Figure 4). For example, the breakage of the salt bridge between the amine group and D138^{3.32} occurred when a water molecule reached the salt bridge, and was often observed to initially form interactions with D138^{3.32}, followed by interacting with the secondary amine, resulting in the breakage of the direct protein-ligand interaction. To appreciate the impact of water molecules on unbinding, we simulated the ligand escape in vacuum and found the maximum value of the rupture force increased to 32 kcal/mol/Å (twice that observed in the solvated systems), suggesting that water-mediated interactions facilitate the ligand's movement along the extracellular cavity (Figure S3).
Movement of antagonists along path B led to the flooding of 15 water molecules into the binding cavity (Figure S4) through the region surrounding the secondary amine of the antagonists.

Path C in β₁AR required the highest force for both ligands. When pulled along this trajectory, both ligands exited through the EL2 in 6–7 simulations with the rupture forces reaching up to 20 kcal/mol/Å. In the case of b₂AR, the ligand escaped along path C without going through EL2 in all trajectories. EL2 has high mobility and therefore tends to cover the extracellular mouth in β₁AR, whereas it has a rigid position and localizes close to the side of helices in b₂AR owing to restriction caused by the extracellular salt bridge. The average root-mean-square deviation values of the β₁AR backbone along three pathways, provided in Figure S5, show higher fluctuation of the receptor in path C, indicating that large structural rearrangements are required to allow the ligand to exit the receptor along this pathway. This is due not only to a steric clash with the hydrophobic tail of K347, but was also a result of the required breakage of the salt bridge between E205EL2 and R351EL3 in several trajectories, which formed and remained stable during equilibration in our cMD simulations.

**Pulling from the β₂ adrenergic receptor**

Simulations of ligand unbinding from b₂AR showed that, contrary to what was observed for β₁AR, path C required the application of the smallest force to bring the antagonists to the surface. This is consistent with recently published work on binding of the β-adrenoblockers to b₂AR using cMD, where the authors showed that ligands bind preferably to the receptor from the extracellular side of helices 5, 6 and 7 (9). In our simulations, we monitored the hydrogen bond network between the side chains of N293EL5, H296EL6, and Y308EL7 (Figure 7A,B), which drives them close to helix 6, enlarges the lateral channel, and thus facilitates the ligand’s movement along path C. To monitor the changes in the size of the lateral channel, we calculated the radius of gyration along path C, as shown in Figure 7C. The hydrogen bond network that exists only during the unbinding process increases the radius of gyration compared to its value in the cMD of the antagonist-receptor complexes (Figure 7D). The dynamics of this network of interactions in the unbinding of ICI-118551 from b₂AR through path C are shown in Movie S2.

In paths A and B, the higher peaks in the force profile appear to be a result of the antagonists breaking the D192EL2-K305EL7 salt bridge at the extracellular part, in addition to the disruption of the salt bridge in the binding site. Along these paths, the antagonists escaped without the breakage of the salt bridge in only seven of 40 trajectories, while the unbinding along pathway C did not require the breakage of the salt bridge in all MD trajectories. The average root-mean-square deviation values of the b₂AR backbone along the three pathways, provided in Figure S5, shows larger deviation of the receptor in paths A and B at the time of the salt bridge breakage, supporting a somewhat obstacle role of the salt bridge.

The sequence of breakage of the original interactions during the unbinding of Esmolol and ICI-118551 was similar to that in β₁AR. Figure 8 shows the time dependence of different ligand-protein interactions along unbinding pathway C. The ester group of Esmolol formed and broke interactions with S204EL5 and occasionally with N293EL5 at the beginning of the unbinding process, while final breakage of this interaction resulted in the first peak in the force profile. Overall, the ester group of Esmolol formed fewer hydrogen bonds along the unbinding pathway in b₂AR than in β₁AR, as shown in Figure 5, while both ligands left the receptor surface by breaking interactions with D300EL3. Our data, as well as the recently published cMD simulations of ligand binding to b₂AR, suggest that D300EL3 likely plays the recognition role of the charged amine of the ligand in a cellular environment, similar to D217EL2 and D356EL3 in β₁AR.
It was also observed that Esmolol escaped quicker than ICI-118551 from β2AR. This is a result of ICI-118551 forming extra aromatic and hydrophobic interactions with Y308^7.35, H296^6.58, F193^EL2 and F194^EL2 through its large hydrophobic moiety along the pathway, as shown in Figure 8. F194^EL2 formed hydrophobic interactions with ICI-118551 and followed the ligand to the solvent environment in several trajectories through movement of its backbone and side chain. The distance between the centre of the aromatic ring of H296^6.58 and antagonists remained 5Å during 250ps in several trajectories, suggesting formation of aromatic interactions. We propose that the side chain of F194 and antagonists remained 5Å during 250ps in several trajectories, suggesting formation of aromatic interactions. We propose that the side chain of F194 and antagonists remained 5Å during 250ps in several trajectories, suggesting formation of aromatic interactions. We propose that the side chain of F194 and antagonists remained 5Å during 250ps in several trajectories, suggesting formation of aromatic interactions.

Within the preferable pathways for ligand unbinding from β1AR and β2AR, we found that the distinct moieties of antagonists have a different pattern of interactions in the receptors. The unbinding of Esmolol is governed through the interchange of hydrogen bonds of the ester group, involving K347^6.58 in β1AR, whereas ICI-118551 does not form interactions with this residue. In contrast, the unbinding from β2AR is likely driven by hydrophobic and aromatic interactions with Y308^7.35, H296^6.58, F193^EL2 and F194^EL2 and, thus, the hydrophobic moiety of ICI-118551 spends more time interacting with these residues than Esmolol. The ester group of Esmolol forms few hydrogen bonds in Y308^7.35, H296^6.58, F193^EL2 and F194^EL2 and, thus, the hydrophobic moiety of ICI-118551 spends more time interacting with these residues than Esmolol.

Side-effects caused by many β-antagonists are owing to their promiscuous binding to closely related subtypes of the adrenoceptors (β1AR and β2AR). Therefore, understanding the molecular basis of β-antagonist selectivity is likely to provide a novel rationale for the discovery of selective ligands. In this work, we have compared the unbinding process of β1- and β2-selective antagonists from β1AR and β2AR by applying steered MD simulations and have shown the potential for a kinetic basis of antagonist selectivity, in addition to the dynamic binding site selectivity caused by different geometry of the conserved N344^6.55. Remarkably, the calculated average rupture force and work profiles from multiple unbinding trajectories suggest that the perpendicular to the membrane direction (path B) is likely the preferable pathway for both ligands to dissociate from β1AR, whereas the lateral direction, composing helices 5, 6 and 7 (path C) is always favourable in β2AR. The calculated ΔPMF of the antagonists for these pathways correlates with the observed ligand selectivity between the receptors. We expect that accuracy of ΔPMF calculations involving hundreds replicates will increase and provide more precise estimation of the ligand binding affinities. However, this approach could be challenging for examining binding activities in the high-throughput scale for many potential ligands.

When pulled in each of the three directions to the extracellular surface, the non-conserved K347^6.58, the E205^EL2-R351^EL3 salt bridge, which is localized in the lateral side in β1AR, and the D193^EL2-K305^EL3 salt bridge, which faces the binding cavity in β2AR, hinder the antagonists from unbinding in the other directions. The relatively equal force profile of path A and B are similar to two favourable unbinding channels in β1AR identified recently by Gonzalez and co-workers (10), while the preference of path C in our short simulations of unbinding from β2AR is in agreement with the recently published 1–19 μseconds conventional MD simulations of ligand binding to β2AR (9).

Within the preferable pathways for ligand unbinding from β1AR and β2AR, we found that the distinct moieties of antagonists have a different pattern of interactions in the receptors. The unbinding of Esmolol is governed through the interchange of hydrogen bonds of the ester group, involving K347^6.58 in β1AR, whereas ICI-118551 does not form interactions with this residue. In contrast, the unbinding from β2AR is likely driven by hydrophobic and aromatic interactions with Y308^7.35, H296^6.58, F193^EL2 and F194^EL2 and, thus, the hydrophobic moiety of ICI-118551 spends more time interacting with these residues than Esmolol. The ester group of Esmolol forms few hydrogen bonds in β2AR during unbinding.
By simulating unbinding from the adrenoceptors, we assume that the binding process takes place in an opposite but a similar fashion; thus, the proposed residues underlying the unbinding likely play important roles during binding as well. The hydrophobic surfaces between alprenolol and Y308, H296, F193, EL2 have been recently observed by Dror and co-workers in long simulations of alprenolol binding (9).

In our simulations, K347 coordinates the antagonist unbinding by restricting the escape from one side of the extracellular surface and utilizing a hydrogen bond to translate the antagonist to the extracellular surface through another route, providing selective features of antagonist unbinding in β1AR. Interestingly, K347 in human β1AR is replaced by asparagine in turkey β1AR, which can play a different role in ligand binding and unbinding, which explains recent simulations of dihydroalprenolol binding that showed the binding pathway was similar to β2AR (9). In addition, in a recent pharmacological study, it was observed that the β1AR-selective antagonist, CGP20712A, in human is not selective in turkey (34). With both species β1AR receptors having identical binding sites, we hypothesize that the residue difference at the extracellular surface is responsible for the different binding profiles of antagonists between these two proteins.

Our study suggests a kinetics basis of antagonist selectivity in the adrenoceptors in which the non-conserved residues at the extracellular surface form selectivity filters that recognize the molecule from the soluble environment by its shape and physicochemical properties, and bring it into the binding site through a cascade of specific intermediate interactions. The mapping of structures taken from the unbinding trajectories by hydrophobic and electrostatic probes highlights differences in the physicochemical properties of the extracellular surface (Figure 9). For example, there is preference for a hydrogen bond acceptor region in β1AR and a hydrophobic region in β2AR, especially in the snapshots where the selective antagonist forms specific interactions during unbinding (Figure 9B–C, E–D). The kinetics-based preferable regions are poorly seen in the ligand-receptor complexes, as shown by the initial structures in Figure 9A,D. Receptor conformations from these unbinding studies can play an important role within a drug design application, for example by designing a ligand with unique binding kinetics by docking molecules or fragments within the binding and unbinding channel, in addition to classical docking to the binding site, or by growing a ligand in such way that it can interact with the extracellular residues forming the preferable maps. Although, there is no experimental proof of kinetic basis of antagonist selectivity in the adrenergic receptors at this moment, our computational study suggests amino acid residues of the receptor extracellular surface for site-directed mutagenesis studies or medicinal chemistry optimization to explore this hypothesis experimentally.

In this work, we have simulated the unbinding event of two antagonists that do not cause conformational changes leading to activation and signalling. The process of agonist binding and unbinding is thought to be more complicated as it involves a series of conformational changes in the receptor structure, resulting in formation of the active state. Fluorescence spectroscopy studies have shown a sequential binding model of agonists in which the partial agonist, dopamine, has rapid binding, whereas a full agonist, noradrenaline, preserves the biphasic binding kinetics in β2-AR (35). We anticipate that there are different interactions in the binding and unbinding pathways of agonists and antagonists. Simulations of the agonist binding process, in which step-by-step conformational changes of the receptor undergoing a transition from the inactive conformation towards the active one, may provide novel molecular features in ligand kinetics, which in turn, could help to interpret ligand efficacy and facilitate design of selective ligands with desired efficacy. The recently published crystal structure of β2AR in complex with the Gs protein, representing the active state, provides the opportunity for a such study (27). Thus, transition from inactive to active states of β2AR in the presence of ligands with different efficacy using adaptive biasing simulations...
recently performed by Provasi and co-workers revealed energetics of the receptor conformational space during activation (36). Transition from active to inactive states in the presence of agonists and atomic details of activation have been recently suggested by Dror and co-workers (37).

Our simulations in a hydrated lipid bilayer at physiological pH were aimed to approximate the realistic in vivo environment. However, GPCRs are often subjected to N- or O-glycosylation within their extracellular regions, which the current computational works on GPCR simulations and our study have neglected. Pharmacological studies have identified that the asparagine residues at positions 6\(N^6\) - the N-terminus of the receptor), 15\(N^15\) and 187\(EL^E\) in \(\beta_2\)AR and at position 15\(N^15\) in \(\beta_1\)AR are N-glycosylated (38,39). The non-conserved N187\(EL^E\) is located in the second extracellular loop faces the extracellular cavity; thus, it may be directly involved in ligand binding through paths C and B in \(\beta_2\)AR. N187\(EL^E\) is replaced by D212\(EL^E\) in \(\beta_1\)AR, without the capability to be glycosylated. It has been shown for several GPCRs that the blockage of glycosylation results in non-functional receptors that are often incapable of ligand binding (40–42). Glycosylation regulates surface expression and dimerization of \(\beta_1\)AR (43). It is not clear how many oligosaccharide chains are present for each glycosylation site in GPCRs; however, it is expected that it could vary from a minimum pentasaccharide structure to a complex oligosaccharide structure containing several oligosaccharide chains (44). Such a 'sweet cloud' confronts the extracellular cavity and likely influences ligand binding and unbinding, providing further structural differences between the receptors. The oligosaccharide cloud can form specific intermediate binding sites, where not only an orthosteric ligand can stick on the way to its binding site, but also an allosteric ligand could interact with this site to modulate receptor activation, enlarging the spectrum of the recently proposed putative allosteric binding sites of \(\beta_1\)AR and \(\beta_2\)AR (45). Importantly, glycosylation produces a stable hydration shell, which causes different entropic cost during ligand binding and unbinding compared to a less ordered hydration shell in the absence of oligosaccharides. Taken together, the next challenge in understanding the molecular basis of ligand binding and unbinding is involvement of receptor active states and glycosylation in GPCR simulations.

**Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

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Figure 1.
Structures of β₁AR-selective Esmolol and β₂AR-selective ICI-118551. Biological activities are taken from references 5 and 7.
Figure 2.
The extracellular side of the adrenergic receptors. (A) the $\beta_1$ adrenergic receptor in the complex with Esmolol, (B) the $\beta_2$ adrenergic receptor in the complex with ICI-118551, (C) the schematic representation of the binding site interactions between Esmolol and the $\beta_1$ adrenergic receptor, and (D) the schematic representation of the binding site interactions between ICI-118551 and the $\beta_2$ adrenergic receptor. The snapshots for the images were taken from the molecular dynamics trajectories. The binding site residues and residues involved in ligand unbinding are visualized in the stick-like representation in A and B. Pulling directions are shown in arrows.
Figure 3.
The average, rupture force and work profiles of antagonist unbinding from the adrenergic receptors in three extracellular pathways. The graphs were plotted against ligand separation from the initial coordinates in the binding site. The unbinding of β₁AR-selective Esmolol and β₂AR-selective ICI-118551 are shown in black and red, respectively.
Figure 4.
Esmolol and ICI-118551 unbinding from the $\beta_1$ adrenergic receptor on the example of one representative trajectory shown in the form of the rupture force, work, hydrogen bonds, hydrophobic and water-mediated contacts projected onto unbinding time and antagonist separation time from the initial position.
Figure 5.
The hydrogen bond profile of the ester group during the Esmolol unbinding from the $\beta_1$ and $\beta_2$ adrenergic receptor.
Figure 6. Evolution of the distance between the centre of the aromatic ring of F218EL and F359 along the unbinding pathway B (A), 30 nseconds of conventional molecular dynamics of the complexes with Esmolol or ICI-118551 (B) and 30 ns of the unoccupied β₁ adrenergic receptor (C).
Figure 7.
The lateral channel to the extracellular surface composing helices 5, 6 and 7. The H296-Y308-N293 hydrogen bond network of interactions in the β2 adrenergic receptor (A, B) along the ligand unbinding trajectories, radius of gyration of the lateral channel, along unbinding pathway C (C) and radius of gyration of the lateral channel in 30 nseconds classical MD of the ligand-receptor complexes (D).
Figure 8.
Esmolol and ICI-118551 unbinding from the β₂ adrenergic receptor on the example of one representative trajectory shown in the form of the rupture force, work, hydrogen bonds, hydrophobic and water-mediated contacts projected onto unbinding time and antagonist separation time from the initial position.
Figure 9.
Hydrophobic and hydrogen bond acceptor grids in the structural snapshots of the $\beta_1$ and $\beta_2$ adrenergic receptors taken from the antagonist unbinding trajectories.
Table 1

The atoms of the receptors and ligands to which an external force has been applied to pull Esmolol and ICI-118551 in three extracellular directions from the β₁ and β₂ adrenergic receptors

<table>
<thead>
<tr>
<th>Path</th>
<th>Cα atom of a residue in β₁/β₂ adrenoceptors</th>
<th>Ligand atom (Esmolol/ICI-118551)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Path A (TM7, TM1 &amp; TM2)</td>
<td>F₅.₄₇ (233/208)</td>
<td>C10/C9</td>
</tr>
<tr>
<td></td>
<td>F₆.₅₂ (341/290)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>S₅.₄₆ (232/207)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>T₃.₃₆ (143/118)</td>
<td></td>
</tr>
<tr>
<td>Path B (Straight)</td>
<td>F₅.₄₇ (233/208)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>M₂.₃₃ (107/82)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>T₃.₃₆ (143/118)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>W₆.₄₄ (337/286)</td>
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</tr>
<tr>
<td>Path C (TM5, TM6 &amp; TM7)</td>
<td>I₃.₄₀ (146/121)</td>
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</tr>
<tr>
<td></td>
<td>M₂.₃₃ (107/82)</td>
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<tr>
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<td>T₃.₃₆ (143/118)</td>
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</tr>
<tr>
<td></td>
<td>Y₇.₄₃ (367/316)</td>
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