Statin use, Candidate Mevalonate Pathway Biomarkers, and Colon Cancer Survival in a Population-Based Cohort Study


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Title
Statin use, candidate mevalonate pathway biomarkers, and colon cancer survival in a population-based cohort study.

Running Title
Statins and colon cancer survival.

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ABSTRACT

Background

Statin use after colorectal cancer diagnosis may improve survival but evidence from observational studies is conflicting. The anti-cancer effect of statins may be restricted to certain molecular subgroups. In this population-based cohort study the interaction between p53 and HMGCR expression, KRAS mutations, and the association between statin use and colon cancer survival was assessed.

Methods

The cohort consisted of 740 stage II and III colon cancer patients diagnosed between 2004-2008. Statin use was determined through clinical note review. Tissue blocks were retrieved to determine immunohistochemical expression of p53 and HMGCR in tissue microarrays and the presence of KRAS mutations in extracted DNA. Cox proportional hazards models were used to calculate hazard ratios (HR) and 95% confidence intervals (CI) for colorectal cancer-specific and overall survival.

Results

Statin use was not associated with improved cancer-specific survival in this cohort (HR=0.91, 95% CI 0.64-1.28). Statin use was also not associated with improved survival when the analyses were stratified by tumour p53 (wild-type HR=1.31, 95% CI 0.67-2.56 versus aberrant HR=0.80, 95% CI 0.52-1.24), HMGCR (HMGCR-high HR=0.69, 95% CI 0.40-1.18 versus HMGCR-low HR=1.10, 95% CI 0.66-1.84), and KRAS (wild-type HR=0.73, 95% CI 0.44-1.19 versus mutant HR=1.21, 95% CI 0.70-2.21) status.
Conclusion

Statin use was not associated with improved survival either independently or when stratified by potential mevalonate pathway biomarkers in this population-based cohort of colon cancer patients.

KEYWORDS

Hydroxymethylglutaryl-CoA reductase inhibitors; Tumour suppressor protein p53; HMGCR protein; KRAS; Colonic neoplasms; Survival.
INTRODUCTION

Statins are commonly prescribed lipid lowering medications that inhibit the enzyme 3-hydroxy-3-methylglutaryl coenzyme-A reductase (HMGCR). (Ng et al, 2011) In addition to their cholesterol lowering action they may have pleiotropic anti-cancer effects through inhibition of the mevalonate pathway. (Bardou et al, 2010; Thurnher et al, 2012) However observational data assessing the association between post-diagnostic statin use and colorectal cancer survival lacks consistency. (Ng et al, 2011; Mace et al, 2013; Cardwell et al, 2014a; Krens et al, 2014; Hoffmeister et al, 2015; Kim et al, 2015; Lim et al, 2015; Zanders et al, 2015) As a robust association has yet to be identified, caution is required in proceeding with clinical trials assessing the role of statins as novel adjuvant agents. In addition, colorectal tumours are known to display significant molecular heterogeneity. (Ogino et al, 2012) A molecular pathological epidemiology approach is therefore required to determine if the potential anti-cancer effect of statins is confined to specific molecular subgroups. (Ogino et al, 2011) Candidate mevalonate pathway biomarkers that may differentiate tumours more likely to respond to statin therapy include HMGCR, p53 and KRAS.

The seminal work by Freed-Pastor et al implicates the mevalonate pathway as a potential therapeutic target for tumours bearing mutations in TP53. (Freed-Pastor et al, 2012) They demonstrated that statins were able to reverse the malignant phenotype of p53 mutant but not p53 wild-type breast cancer cells in vitro. Similarly, an in vivo breast cancer study demonstrated that the anti-proliferative effect of statins was limited to tumours that overexpressed HMGCR. (Bjarnadottir et al, 2013) Finally, RAS signaling may be inhibited by statin-induced depletion of downstream isoprenoids required for posttranslational prenylation of small GTPases.
like ras and rho. (Bardou et al., 2010; Ng et al., 2011; Thurnher et al., 2012) Prenylation of k-ras makes the protein lipophilic and ensures translocation to the cell membrane where it can exert its proliferative effects. (Konstantinopoulos et al., 2007; Krens et al., 2014) Based on this hypothesis the effect of statins on colorectal cancer survival may differ according to KRAS gene mutation status. (Ng et al., 2011) The aim of this study was therefore to assess the interaction between statin use, the potential mevalonate pathway biomarkers p53, HMGCR and KRAS, and survival in a population-based cohort study of patients with stage II and III colon cancer.
METHODS

Study cohort

The Northern Ireland Cancer Registry was used to identify 1,426 stage II and III colon cancer patients undergoing surgical resection between 2004 and 2008 (Figure 1). Rectal cancers were excluded as neoadjuvant radiotherapy could potentially alter tumour expression profiles. Ethical approval through the Northern Ireland Biobank (NIB ref. 13-0087) permitted retrieval of formalin fixed, paraffin-embedded (FFPE) tissue blocks for patients within two of the five regional Health and Social Care trusts. For this molecular pathological epidemiology study, the final cohort was subsequently restricted to only include patients within the biobank remit (n=740, 51.9%). These patients were representative of the overall Northern Ireland cohort with respect to age, sex, stage and adjuvant chemotherapy receipt (Supplementary Table 1).

Clinicopathological variables and follow-up

The Clinical Oncology Information System (COIS), a prospective electronic record of cancer patient management, was used to collect clinical variables including adjuvant chemotherapy use, prescription medication use, family history of colorectal cancer and Eastern Cooperative Oncology Group (ECOG) performance status. This process was supplemented by a manual chart review when insufficient information was recorded on COIS or no record was present. Pathological variables were retrieved from full pathology reports. Occurrence and cause of death were assessed via data linkage to the Northern Ireland Registrar General’s Office (follow-up censored 31st December 2013). Colorectal cancer-specific deaths were defined as those with an underlying cause of death International Classification of Disease code C18, C19, C20 (anus) and/or C26 (other and ill-defined digestive organs).
**Drug exposure assessment**

Statin exposure (user versus non-user) based on current prescription medications was assessed at a single perioperative time point for all patients. When medication information was available on COIS this time point was the initial post-operative oncology review. When medication information was not available on COIS statin exposure was determined from the post-operative hospital discharge letter. Information on medication dosage was not consistently recorded on COIS and therefore not considered. Information on aspirin exposure was also assessed using these methods. Our research group has previously demonstrated that 98.5% of aspirin prescriptions after colorectal cancer diagnosis in the United Kingdom (UK) are for low-dose (75mg) aspirin. (Cardwell *et al*, 2014b) Aspirin exposure in this study is therefore considered representative of low-dose aspirin.

**Tumour molecular analysis**

FFPE blocks were retrieved for 89.3% of the cohort (661 of 740). Three 1.0mm diameter tissue cores were extracted from representative areas within donor blocks and inserted into recipient blocks using a manual tissue arrayer (Estigen, Tartu, Estonia) as described previously. (Zhang *et al*, 2003; Boyle *et al*, 2014) The immunohistochemistry methods for p53 (DO-7 antibody clone to p53, Dako UK Ltd, Ely, UK - catalogue number M7001) and HMGCR (Atlas Antibodies AB, Stockholm, Sweden - catalogue number HPA008338) staining are described in the Supplementary Methods. *QuPath* (Queen’s University Belfast, Northern Ireland) image analysis software facilitated digital immunoscroing (Supplementary Methods). A H-score was calculated based on the extent and intensity of cytoplasmic or nuclear
staining where appropriate (H-score = 3 x % of strongly staining cytoplasm + 2 x % of moderately staining cytoplasm + 1 x % of weakly staining cytoplasm, giving a range of 0 to 300). (McCarty et al, 1986)

A three-tier scoring system was applied to differentiate normal (non-extreme) from aberrant (extreme positive or extreme negative) patterns of p53 expression (Figure 2). Cores were designated as extreme negative if there was confluent negative staining within the represented population of tumour nuclei. Diffuse strong positivity was considered representative of extreme positive expression and intermediate heterogeneous expression was considered a non-extreme (normal) pattern of staining. (Boyle et al, 2014) Selection and validation of the p53 cut points are described in the Supplementary Methods. HMGCR H-scores were categorised into tertiles for prognostic analyses and dichotomised around the median value for survival analyses that tested the interaction between statin use and HMGCR expression. Representative images of HMGCR expression are shown in Supplementary Figure 1. Based on these methods the final p53 and HMGCR categories were determined using the median of the three available H-scores for each case.

Detailed methods for DNA extraction, KRAS mutation analysis and microsatellite instability (MSI) status using commercially available kits are provided as Supplementary Methods. Briefly KRAS mutation status was assessed using previously described methods for the ColoCarta Panel (Agena Bioscience, Hamburg, Germany) (Fumagalli et al, 2010) while MSI status was determined using five mononucleotide repeat markers (BAT-25, BAT-26, NR-21, NR-24 and MONO-27).

**Statistical analysis**
All statistical analysis was performed using Stata 13 (StataCorp, College Station, TX, USA). The chi-square test was used to compare characteristics between statin users and non-users. The primary outcome of this study was colorectal cancer-specific survival and the secondary outcome was overall survival. The association between statin use and survival was assessed in the whole cohort and then in analyses stratified by biomarker status. Only cases with information on statin exposure (known user versus known non-user) were included in the former analysis. Only cases with available exposure information and tissue for biomarker assessment were included in the subsequent stratified analyses. Other missing categorical data were coded as unknown.

Survival analysis was performed using the Cox proportional hazards model to calculate hazard ratios (HRs) and associated 95% confidence intervals. The multivariable models for colorectal cancer-specific survival adjusted for age, gender, year of diagnosis, grade, MSI status, Eastern Cooperative Oncology Group performance status, family history of colorectal cancer, adjuvant chemotherapy use, stage and aspirin use. In addition to these variables, the overall survival multivariable model also adjusted for Charlson Comorbidity Index score as a continuous variable. Analyses were stratified by biomarker status. Interaction terms for statin use and p53, HMGCR, or KRAS were then included in the Cox model and the Wald test was used to assess for statistical interaction. Sensitivity analysis was performed using complete-case data (cases with missing data were excluded). All P values were two-sided and a value <0.05 was considered statistically significant.
RESULTS

Patients

Information on prescription medication use was available for 91.9% (n=680) of patients in this population-based cohort study. Overall, 25.3% (n=172) used statins. Compared to those with available information on medication use, patients with no information on medication use were older and more likely to be diagnosed in the earlier years of the study. However, there was no difference in stage, grade of tumour differentiation or MSI status (Supplementary Table 2). After a mean follow-up of 5.7 years (range 0-10) there were 299 all-cause and 212 colorectal cancer-specific deaths among these patients.

Statin use and survival

Table 1 summarises the baseline characteristics between statin users and non-users. Statin users were more likely to be older, male, and diagnosed later in the cohort compared to statin non-users. Statin users were also more likely to concomitantly use aspirin. There were a smaller proportion of MSI-high tumours among statin users but the proportion of patients with unknown MSI status was higher in statin non-users. However there was no difference in tumour differentiation grade, stage or the proportion of patients receiving adjuvant chemotherapy between users and non-users. There was also no difference in the proportion of right-sided tumours in statin users compared to non-users (58.1 versus 56.1%).

Statin use at the time of diagnosis was not associated with a significant reduction in colorectal cancer-specific (adjusted HR=0.91, 95% CI 0.64-1.28) or overall mortality (adjusted HR=0.83, 95% CI 0.61-1.12) compared to non-use (Table 2).
**Immunohistochemical expression of p53**

In total n=361 (59.9%) tumours demonstrated an aberrant pattern of p53 immunostaining (extreme positive or extreme negative). The proportion of tumours with aberrant patterns of p53 immunostaining was similar between statin users and statin non-users (59.1% versus 62.0%, P=0.52). Compared to wild-type p53 expression, aberrant p53 immunostaining was associated with a 53% increase in hazard for unadjusted colorectal cancer-specific mortality (HR=1.53, 95% CI 1.13-2.09). However, this association was attenuated when potential confounding variables were included in the multivariable model (adjusted HR=1.38, 95% CI 0.97-1.95, Table 2).

In stratified analysis there was no evidence that the association between statin use and colorectal cancer-specific survival differed by p53 immunostaining patterns (wild-type adjusted HR=1.31, 95% CI 0.67-2.56 versus aberrant adjusted HR=0.80, 95% CI 0.52-1.24). Similar results were observed for overall survival (Table 3).

**HMGCR immunohistochemical expression**

Statin users were more likely to have tumours in the highest HMGCR tertile compared to non-users (43.7% versus 30.8%, P=0.01). As shown in Table 2 however, there was no evidence that higher levels of HMGCR expression were associated with significantly better colorectal cancer-specific or overall survival (adjusted P for trend=0.18 and 0.12 respectively).

There was no evidence of significant associations with colorectal cancer-specific survival among statin users compared to non-users when the cohort was
stratified by tumour HMGCR expression level, although the direction of hazard ratios did differ (HMGCR-high adjusted HR=0.69, 95% CI 0.40-1.18, P=0.17 versus HMGCR-low adjusted HR=1.10, 95% CI 0.66-1.84). Similarly, there was no evidence of a differential benefit for overall survival in statin users compared to non-users when the cohort was stratified by HMGCR status (Table 3).

**KRAS mutations**

KRAS mutation status (mutant versus wild-type) was available for 99.2% of these samples with extracted DNA (594 of 599). Statin users had slightly less KRAS mutant tumours compared to statin non-users although the difference was not statistically significant (34.0% versus 40.2%, P=0.17). Compared to wild-type KRAS, the presence of a KRAS mutation was not associated with significantly worse colorectal cancer-specific survival (adjusted HR=1.12, 95% CI 0.82-1.53, Table 2).

There was no evidence of an improvement in colorectal cancer-specific survival among statin users compared to non-users when the cohort was stratified by KRAS mutation status, although again the direction of effect differed between wild-type and mutant KRAS tumours (KRAS wild-type adjusted HR=0.73, 95% CI 0.44-1.19 versus KRAS mutant tumour adjusted HR=1.21, 95% CI 0.70-2.21). Similar non-significant results were observed for overall survival (Table 3).

**Sensitivity analysis**

In general the associations described above were not markedly altered when a complete case dataset (n=372) was used (Table 4). There was some evidence though that statin use was associated with improved colorectal cancer-specific survival in tumours that had higher levels of HMGCR expression (HMGCR-high
adjusted HR=0.51, 95% CI 0.26-0.97, P=0.04 versus HMGCR-low adjusted
HR=0.92, 95% CI 0.45-1.87, P for interaction=0.05). Previously observed null
associations by KRAS status and p53 expression remained. All associations
became attenuated in analyses evaluating overall survival.
DISCUSSION

In this population-based cohort study of stage II and III colon cancer, perioperative statin prescription was not associated with significantly improved colorectal cancer-specific survival. Similarly, statin use was not associated with better colorectal cancer-specific or overall survival when the cohort was stratified by tumour biomarkers related to the mevalonate pathway.

The hazard ratio for the association between statin use and cancer-specific survival in this cohort was similar to that reported in a recent meta-analysis of colorectal cancer observational studies.(Gray et al, 2016) The pooled estimate from four studies assessing post-diagnostic statin use was non-significant despite including over 19,000 patients. It also suggests that the effect of any association is only likely to be moderate at best (pooled HR=0.84, 95% CI 0.68-1.04). Despite optimistic pre-clinical data these findings confirm the need to evaluate biomarkers that may identify tumours more likely to respond to the potential anti-cancer effects of statins.

This is the first study to assess the interaction between statin use and HMGCR expression in colon cancer. Statin users had a higher proportion of tumours in the highest HMGCR tertile although statin-induced inhibition of the mevalonate pathway is known to trigger a marked increase in the production of inactive HMGCR in vitro.(Goldstein & Brown, 1990; Bengtsson et al, 2014) In the main analysis there was no evidence that statin users had better survival compared to non-users in tumours with higher levels of HMGCR expression. However, in the complete-case subgroup analysis, which excluded cases with any missing data, statin use was associated with better cancer-specific survival in tumours with higher levels of HMGCR expression. This result should be interpreted with caution though, as
multiple hypotheses were tested. Further exploration in additional molecular pathological epidemiology cohorts should be considered, as the complete-case subgroup analysis results are in line with an in vivo breast cancer study which suggests that statins may have an anti-proliferative effect in tumours that overexpress HMGCR. (Bjarnadottir et al, 2013)

Overexpression of HMGCR has been proposed to be prognostic in a number of malignancies including breast (Borgquist et al, 2008; Brennan et al, 2011) and epithelial ovarian cancer. (Brennan et al, 2010) However, a recent population-based breast cancer cohort study failed to demonstrate that overexpression of HMGCR was associated with better survival. (Gustbée et al, 2015) Similarly, overexpression of HMGCR was not associated with improved survival after adjusting for confounding variables in colorectal cancer cases within the Malmö Diet and Cancer Study. (Bengtsson et al, 2014) The present study largely corroborates this finding.

To the best of the authors’ knowledge this is the first study to assess the interaction between statin use, p53 expression and survival in patients with colon cancer. A significant interaction was not identified in this instance but further work is required as TP53 mutation status was not directly assessed. Missense TP53 mutations result in stabilization of an inactive form of p53 resulting in nuclear accumulation and a correlation with the aberrant positive pattern of expression. (Kaye et al, 2010; McCluggage et al, 2011) Only more recently has it been widely appreciated that the aberrant negative pattern of p53 staining is a distinct entity and not part of the spectrum of wild-type staining. (Boyle et al, 2014) This pattern of staining may be attributed to a null TP53 mutation resulting in complete absence of the detectable protein. (Köbel et al, 2010) Importantly though, these patterns should be viewed as a spectrum of functional protein status rather than as a surrogate for
TP53 mutation status as epigenetic silencing may also contribute to aberrant negative expression. (Kaye et al, 2010; Boyle et al, 2014) On this basis, mevalonate pathway gene-expression upregulation associated with mutant p53 (Freed-Pastor et al, 2012) may be specific to mutations of TP53 rather than to alternate circumstances resulting in aberrant expression of the protein. Future studies should therefore assess the interaction between the presence of TP53 mutations, statin use and colon cancer survival before excluding the potential relevance of this biomarker.

Finally, statin use was not associated with improved colorectal cancer-specific or overall survival when the cohort was stratified by KRAS mutation status. This finding is consistent with results from a cohort of 394 patients enrolled in a chemotherapy clinical trial (CALGB 89803) (Ng et al, 2011) and 1209 patients within a German population-based colorectal cancer cohort. (Hoffmeister et al, 2015) Similarly, statin use was not associated with improved progression free survival in cetuximab treated metastatic colorectal cancer patients within the CAIRO2 trial, irrespective of KRAS mutation status. (Krens et al, 2014) The median progression free survival was also similar between the statin and placebo arms of a subgroup of 83 patients with KRAS mutant tumours in a randomized controlled trial of XELIRI/FOLFIRI +/- simvastatin in patients with metastatic colorectal cancer. (Lim et al, 2015)

A major strength of this study is the inclusion of population-representative colon cancer patients. Application of a precise, automated and validated digital immunoscopying system also ensures robust immunohistochemical data that is highly reproducible. As with all observational studies however there may be residual confounding that we were not able to control. A more specific limitation is that data on statin prescription was also only available at a single perioperative time point and
this may not reflect changes in post-diagnostic use. (Paleari et al, 2016) However, a similar European colorectal cancer cohort demonstrated 88% concordance between baseline and long-term statin use. (Hoffmeister et al, 2015) Statin use at this time could also alter tumour behavior as it has previously been reported that pre-diagnostic statin users were less likely to develop KRAS wild-type tumours. (Lee et al, 2011) In the current study the opposite association (non-significant) was observed with a lower proportion of KRAS mutant tumours amongst statin users. Importantly though, assessing medication use at a fixed time point excludes immortal time bias. (Lévesque et al, 2010) The assessment of Ras status was limited to exons 2 and 3 of KRAS in this study. Misclassification could occur for other mutations of KRAS or NRAS although overall these mutations are uncommon (<3%) and it is unlikely that this would greatly alter the stratified analysis.

A further limitation is that information on the type and dose of statin prescribed was not available. It has previously been hypothesised that the potential anti-cancer effect of statins is restricted to lipophilic statins. (Ahern et al, 2014) Also, the serum statin concentrations achieved with cardiovascular protective doses of the medication (e.g. simvastatin 40mg) may not be sufficient to induce the anti-cancer effects observed in preclinical studies. (Lim et al, 2015) Finally, despite being population-based this study lacks power to definitively investigate the interaction between the proposed mevalonate pathway biomarkers, statin use and colon cancer survival.

In summary, statin use was not associated with better survival in this population-based colon cancer cohort study. In keeping with previous studies a survival benefit for statin use was not apparent after stratification by tumour KRAS mutation status. Similar results were also observed for p53 immunohistochemical
status but additional studies should assess TP53 mutation status as a potential biomarker. There was some evidence of a difference in association between statin use and colon cancer survival by tumour HMGCR expression. In general though, this finding was inconsistent and requires further investigation in additional large studies.
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Disclosures

Peter W. Hamilton is Founder and Director in PathXL Ltd. Manuel Salto-Tellez is a senior advisor to PathXL. The authors declare no other conflicts of interest.
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FIGURE LEGEND

**Figure 1.** Selection of stage II and III colon cancer (adenocarcinoma) patients and samples. Abbreviation: ICD – International Classification of Disease; NIB – Northern Ireland Biobank.

**Figure 2.** p53 immunohistochemistry in colon cancer tissue microarrays and associated markup for digital immunoscopying using QuPath image analysis software. Detected cells are color-coded according to their classification: green = non-tumour; blue = negatively staining tumour; yellow = weakly staining tumour; orange = moderately staining tumour; red = strongly staining tumour. (A) Original core from a tumour demonstrating aberrant negative p53 immunostaining. (B) Original core from a tumour demonstrating non-extreme (normal) p53 immunostaining. (C) Original core from a tumour demonstrating aberrant positive p53 immunostaining. (D) QuPath cellular markup in the aberrant negative p53 core. (E) QuPath cellular markup in the non-extreme (normal) p53 core. (F) QuPath cellular markup in the aberrant positive p53 core.