Mutational spectrum of the ZEB1 gene in corneal dystrophies supports a genotype-phenotype correlation


Published in:
Investigative Opthalmology and Visual Science

Document Version:
Publisher's PDF, also known as Version of record

Queen's University Belfast - Research Portal:
Link to publication record in Queen's University Belfast Research Portal

Publisher rights
Copyright 2013 The Association for Research in Vision and Ophthalmology, Inc.
This work is made available online in accordance with the publisher's policies. Please refer to any applicable terms of use of the publisher.

General rights
Copyright for the publications made accessible via the Queen's University Belfast Research Portal is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The Research Portal is Queen's institutional repository that provides access to Queen's research output. Every effort has been made to ensure that content in the Research Portal does not infringe any person's rights, or applicable UK laws. If you discover content in the Research Portal that you believe breaches copyright or violates any law, please contact openaccess@qub.ac.uk.

Download date: 23. Dec. 2018
Mutational Spectrum of the ZEB1 Gene in Corneal Dystrophies Supports a Genotype–Phenotype Correlation

Judith Lechner,1 Durga P. Dash,1 Dorota Muszynska,1 Mohsen Hosseini,2 Fani Segev,3 Sonia George,4 David G. Frazer,4 Jonathan E. Moore,4 Stephen B. Kaye,5 Terri Young,6 David A. Simpson,1 Amanda J. Churchill,7 Elise Héon,2 and Colin E. Willoughby1,4,8

1Centre for Vision and Vascular Science, Queen’s University Belfast, Belfast, Northern Ireland, United Kingdom
2Department of Ophthalmology and Vision Sciences, The Hospital for Sick Children, Toronto, Ontario, Canada
3Meir Medical Center, Kfar Saba, Israel
4Department of Ophthalmology, Belfast Health and Social Care Trust, Belfast, Northern Ireland, United Kingdom
5St. Paul’s Eye Unit, Royal Liverpool University Hospital, Liverpool, United Kingdom
6Center for Human Genetics, Duke University Medical Center, Durham, North Carolina
7School of Clinical Sciences, University of Bristol and Bristol Eye Hospital, Bristol, United Kingdom
8Department of Eye and Vision Science, Institute of Ageing and Chronic Disease, University of Liverpool, Liverpool, United Kingdom

Correspondence: Colin E. Willoughby, Department of Eye and Vision Science, Institute of Ageing and Chronic Disease, Faculty of Health and Life Sciences, University of Liverpool, 3rd Floor, UCD Building, Daleby Street, Liverpool, L69 3GA, UK; drcolinwilloughby@googlemail.com.

JL, DPD, and DM contributed equally to the work presented here and therefore should be regarded as equivalent authors.

Submitted: January 30, 2013
Accepted: April 4, 2013

PURPOSE. Mutations in ZEB1 have been reported in posterior polymorphous corneal dystrophy (PPCD3; MIM #609141) and Fuchs’ endothelial corneal dystrophy (FECD6; MIM #613270). Although PPCD and keratoconus are clinically and pathologically distinct, PPCD has been associated with keratoconus, suggesting a common genetic basis. The purpose of our study was to perform mutational screening of the ZEB1 gene in patients affected with keratoconus or PPCD.

METHODS. Sanger sequencing of ZEB1 was performed in 70 unrelated patients with keratoconus and 18 unrelated patients with PPCD. Real-time quantitative PCR (RT-qPCR) was performed on RNA from cultured corneal keratocytes obtained from a keratoconic patient harboring a missense ZEB1 mutation (p.Gln640His) undergoing corneal transplantation.

RESULTS. Mutational analysis of ZEB1 in PPCD identified a previously reported frameshift mutation (c.1578_1579insG) and a novel nonsense mutation (c.2249C>T) in exon 7 of ZEB1 causing the insertion of a stop codon: p.Ser750X. In the keratoconus cohort, a novel heterozygous pathogenic mutation in exon 7 (c.1920G>A; p.Gln640His) of ZEB1 was identified in a family affected with keratoconus and Fuchs’ endothelial corneal dystrophy. RT-qPCR performed on cultured corneal keratocytes harboring the missense ZEB1 mutation (p.Gln640His) demonstrated that COL4A1 and COL4A2 were markedly downregulated, and COL4A3, COL4A4, and COL8A2 were moderately downregulated.

CONCLUSIONS. Our data combined with the previously reported mutational spectrum of ZEB1 support the idea of a genotype–phenotype correlation: missense substitutions in the ZEB1 protein are associated with keratoconus and PPCD, whereas protein truncating ZEB1 mutations result in PPCD. The dysregulation of z-type IV collagens represents a common link between ZEB1 mutation and the clinical phenotypes (PPCD, FECD, and keratoconus).

Keywords: keratoconus, posterior polymorphous corneal dystrophy, Fuchs’ endothelial dystrophy, ZEB1, corneal dystrophies, hereditary

The corneal dystrophies represent a group of inherited corneal disorders that are genetically and phenotypically heterogeneous, and have been classified by phenotypic description, pathologic examination, and genetic analysis.1 Posterior polymorphous corneal dystrophy (PPCD) is a rare autosomal dominant disorder of the corneal endothelium1 that shows highly variable expressivity in age of presentation and clinical severity.2–4 Clinically, PPCD is characterized by the presence of vesicles, bands, and geographic opacities involving the endothelium and Descemet’s membrane.5,6 The inter- and intrafamilial phenotypic spectrum of PPCD is wide, with some patients remaining asymptomatic with a nonprogressive course, while others show severe progressive disease developing visual impairment due to corneal edema and requiring corneal transplantation.2,4–6

PPCD has been mapped to three different genetic loci,5,7,8 and there is evidence of further genetic heterogeneity9–11 PPCD1 (MIM 605020) was mapped to chromosome 20p11.2-q11.2, and mutations were reported in visual system homeobox gene 1 (VSX1; MIM 605020) in PPCD and keratoconus (KTNC1; MIM 148300).12 PPCD2 (MIM 609140) results from mutations in COL8A2 encoding the alpha 2 chain of type VIII collagen (MIM 120252), located on chromosome 1p34.3-p32.38. Mutations in COL8A2 also have been reported in Fuchs’ endothelial
corneal dystrophy (FEDC1; MIM 136800). PPCD3 (MIM 609141) was mapped to chromosome 10p11.2 and mutations were detected in zinc finger E box-binding homeobox 1 (ZEB1; MIM 189909) which also is known as transcription factor 8 (TCFP8) by some investigators.5–11 Mutations in ZEB1 account for approximately 30% of PPCD patients.5–9,11 and all the reported ZEB1 mutations responsible for PPCD3 are trunctating.5,9–11 Missense mutations in ZEB1 have also been reported in Fuchs’ endothelial corneal dystrophy (FEDC6, MIM 613270).14,15

Although PPCD, Fuchs’ endothelial dystrophy, and keratoconus are clinically and pathologically distinct, keratoconus has been associated with PPCD.16–20 and Fuchs’ endothelial dystrophy,10 suggesting a common link.12,17–21 Here, we report the results of ZEB1 sequencing in a cohort of patients with keratoconus and PPCD. Our data combined with the previously reported mutational spectrum of ZEB1 supports a genotype-phenotype correlation, in which protein truncating ZEB1 mutations result in PPCD, and missense substitutions in the ZEB1 protein are associated with keratoconus and Fuchs’ endothelial dystrophy.

METHODS
Clinical Assessment

Patients affected with either keratoconus or posterior polymorphous corneal dystrophy were recruited as part of ongoing studies from Belfast (Belfast Health and Social Care Trust, Belfast, UK), Bristol (Bristol Eye Hospital, Bristol, UK), and Toronto (Department of Ophthalmology and Vision Sciences, The Hospital for Sick Children, University of Toronto, Toronto, Ontario, Canada). A total of 96 unrelated individuals (192 chromosomes) without ocular disease (aged 60 and over) was used as normal controls. All studies adhered to the tenets of the Declaration of Helsinki, and were approved by the relevant institutions with all participants giving written informed consent. In subjects with pathogenic sequence variants a medical history was taken to identify nonocular features previously associated with ZEB1: inguinal hernias, hydroleces, Dupuytren’s contractures, bony lumps, spinal disease, otosclerosis, and Osgood-Schlatter disease.5,11

Patients were diagnosed with PPCD if they exhibited the characteristic endothelial changes in both eyes (vesicles, bands, or geographic opacities in the endothelium) on slit-lamp biomicroscopy. The diagnosis of keratoconus was based on well-established clinical signs on slit-lamp biomicroscopy and cycloplegic retinoscopy, and a confirmatory videokeratographic map.22,23 Videokeratographic evaluation was performed on each eye using the Topographic Modelling System-1 (Computed Anatomy, Inc., New York, NY), Orbscan II (Bausch & Lomb, Salt Lake City, UT), or the Pentacam (Oculus, Wetzlar, Germany). When determining the familial segregation of ZEB1 sequence variants, all available family members underwent clinical and topographic examination to determine their clinical status and were screened molecularly.

DNA Extraction and Sequencing

Genomic DNA was extracted from peripheral blood leukocytes using a Wizard Genomic DNA Purification Kit (Promega, Southampton, UK) according to the manufacturer’s instructions. PCR primers for amplification and sequencing of the exons, flanking intron sequences, and 5’ and 3’ untranslated regions (UTR) of ZEB1 were identical to those described by Krafchak et al.3 with adapted conditions, except for two additional primers for sequencing that were designed using Primer3 (v. 0.4.0) software (available in the public domain at http://frodou.wi.mit.edu/primer3/)4 and are listed in Supplementary Table S1. Bidirectional cycle sequencing was performed using BigDye Terminator v3.1 cycle sequencing kit (Applied Biosystems, Warrington, UK) and electrophoresed on an ABI PRISM 3730 DNA sequencer (Applied Biosystems, Warrington, UK). Sequencing results were analyzed manually using the sequence analysis software SeqScape 2.1.1 (Applied Biosystems). Identified sequence variants were described according to the guidelines published by the Human Genome Variation Society. Variants were annotated in accordance with NCBI NM_030751.5 (Build GRCh37/hg19).

Deleterious structural effects of amino acid substitutions on protein function were assessed using the PolyPhen-2 (Polymorphism Phenotyping v2; available in the public domain at http://genetics.bwh.harvard.edu/pph2/),25 Sorting Intolerant from Tolerant (SIFT; available in the public domain at http://sift.jcvi.org/),26 and Pathological Mutations (PMut; available in the public domain at http://mmb2.pcb.ub.es:8080/PMut/)27 algorithms. The conservation of the affected amino acid across species was analyzed using Homologene (available in the public domain at http://www.ncbi.nlm.nih.gov/homologene/) and multiple sequence alignment using the UCSC Genome Browser (available in the public domain at http://genome.ucsc.edu/). ZEB1 sequence variants that were absent from controls were assessed to determine potential effects on splicing using Human Splicing Finder Version 2.4.1 (available in the public domain at http://www.umd.be/HSF/).28

Human Corneal Keratocyte Cell Culture

Human corneal keratocytes were cultured directly from residual normal donor corneoscleral rims (n = 4) and from corneal tissue obtained from a patient harboring a missense mutation in ZEB1 (c.1920G > T; p.Gln640His) undergoing corneal transplantation. Corneal stromal tissue was dissected, fragmented, and plated on 60 mm Petri dishes. The cells were maintained separately in Dulbecco’s modified Eagle’s medium/nutrient Ham’s F-12 (1:1) medium (Gibco; Invitrogen Ltd., Paisley, UK) supplemented with 10% fetal calf serum (FCS) and penicillin/streptomycin mix (1:1) (Sigma-Aldrich Company Ltd., Dorset, UK), at 37°C in a 5% CO2 incubator (MiniGalaxy A C&M Scientific; Scientific Laboratory Supplies Ltd., Nottingham, UK). The phenotype of the cells was assessed on the basis of the distinctive morphology of human corneal keratocytes and positive vimentin (DAKO Denmark A/S, Glostrup, Denmark), and CD34 (Santa Cruz Biotechnology, Inc., Santa Cruz, CA) immunofluorescence staining. The cultured cells also were negative for cytokeratin 3 (Millipore, Billerica, MA) immunofluorescence indicating an absence of corneal epithelial cell contamination. Cells were cultured until 90% confluence (10–14 days) and total RNA was extracted using the TRI Reagent (Sigma-Aldrich Company, Ltd.) and quantified on the Nanodrop-1000 (Nanodrop Technologies, Wilmington, DE). RNA quality was determined with the Bioanalyzer (Agilent Technologies UK Limited, Cheshire, UK).

Real-Time Quantitative PCR (RT-qPCR)

Total RNA (1 µg) isolated from human corneal keratocytes was reverse transcribed into cDNA using the QuantiTect Reverse Transcription Kit (Qiagen, West Sussex, UK) according to manufacturer’s protocol. RT-qPCR was performed by using a real-time PCR system (LightCycler 480; Roche Diagnostics, Basel, Switzerland). Primer sequences were obtained from PrimerBank (available in the public domain at http://pga.mgh.harvard.edu/primerbank/)29 for COL4A1, COL4A2, COL4A3, COL4A4, COL4A5, COL4A6, COL8A2, and ZEB1. Real-time PCR reactions were performed in 10 µl final volume
containing 10 ng of cDNA, gene-specific primers, and fluorescent dye SYBR green I (Fermentas UK Ltd., Cambridge, UK; conditions available on request). All PCR reactions were performed in technical triplicates. All mRNA levels were measured as CT threshold levels and were normalized with reference genes30; glyceraldehyde-3-phosphate dehydrogenase (GAPDH), peptidylprolyl isomerase A (PPIA), ubiquitin C (UBC), hypoxanthine quinine phosphoribosyltransferase 1 (HPR1), and ribosomal protein L5 (RPL5) CT values. Values were expressed as fold increase over the corresponding values for control by the 2^{-ΔΔCT} method. The ΔCt validation experiments showed similar amplification efficiency for all templates used (difference between linear slopes for all templates less than 0.1). Three independent experiments were performed and the average (± SEM) results were calculated as CT threshold levels and were normalized with GAPDH. The mRNA levels were measured as CT threshold levels and were normalized with GAPDH.

### Results

Sanger sequencing of the entire coding region of ZEB1 (exons 1–9) was performed on 70 unrelated patients with keratoconus (21 familial and 49 sporadic cases) and 18 unrelated patients with PPCD (7 familial and 11 sporadic cases). Pathogenicity of identified sequence variants was determined by filtering using the in-house sequencing data from 96 control subjects, data from dbSNP (Build 137), the May 2012 release of the 1000 Genomes (1 KG) Project (available in the public domain at http://browser.1000genomes.org/index.html), and the Exome Variant Server (EVS), NHLBI Exome Sequencing Project (ESP; Seattle, WA, available in the public domain at http://evs.gs.washington.edu/EVS/), segregation in familial cases, residue conservation, splicing analysis, and structural modeling.

### Mutational Analysis of ZEB1 in Keratoconus

In patients with keratoconus, we identified 7 heterozygous sequence variants; 3 nonsynonymous and 4 synonymous (Table 2). Three of these were known SNPs (rs80194531, rs7918614, and rs55238902) and one was a novel SNP (c.5177A > T; p.Pro1059Pro) seen in 5% (5/96) of controls, although absent from the 1KG and EVS datasets. There were 3 novel heterozygous sequence variants (c.1574G > A; p.Gly525-Glu, c.1920G > T; p.Gln640His, and c.2673G > C; p.Pro891-Pro), which were absent from controls (192 chromosomes), and the 1KG and ESP datasets.

The novel heterozygous sequence variant exon 7, c.1574G > A (p.Gly525Glu), was identified in a patient with sporadic keratoconus. This sequence variant results in a nonconservative substitution of a nonpolar neutral amino acid (glycine) with a polar negatively charged hydrophilic residue (glutamic acid). The glycine at position 525 is well conserved in primates, although in the chicken and shrew the residue is a glutamic acid. PMut predicted that the variant was pathologic with a score of 0.8 and high reliability, whereas PolyPhen-2 and SIFT predicted this substitution was benign and tolerated, respectively. Given this evidence, we proposed that p.Gly525-Glu is classified as potentially pathogenic. As the novel synonymous change c.2673G > C (p.Pro891Pro) was absent from controls, and the 1KG and ESP datasets, we sought to determine whether there was an impact on splicing due to the impact of the nucleotide substitution on exonic splicing enhancers (ESE) and silencers (ESS).31 On ESE analysis the nucleotide change c.2673G > C alters the predicted ESE binding site, but we were unable to obtain RNA from the patient to assess whether this nucleotide change results in aberrant ZEB1 splicing.

The novel heterozygous pathogenic mutation in exon 7, c.1920G > T (p.Gln640His), was identified in two siblings with keratoconus (Fig. 2B). The nucleotide change causes the conserved polar amino acid glutamine to be substituted by the positively charged, basic amino acid histidine. The glutamine at position 640 is the last amino acid of the homeodomain, and is well conserved across primates and other vertebrates as demonstrated in Figure 2C. SIFT analysis predicted that the variant was damaging, whereas PolyPhen-2 and PMut predicted this substitution was benign and neutral, respectively.

---

**Table 1. ZEB1 Sequence Variants Identified in the Posterior Polymorphous Corneal Dystrophy Patient Cohort**

<table>
<thead>
<tr>
<th>Nucleotide Position</th>
<th>Amino Acid Change</th>
<th>Exon/Intron</th>
<th>Cases</th>
<th>Controls</th>
<th>dbsNP rs #</th>
<th>1KG (MAF %)</th>
<th>ESP (MAF %)</th>
<th>Classification</th>
</tr>
</thead>
<tbody>
<tr>
<td>c.192C &gt; T</td>
<td>p.Asp61ASP</td>
<td>Exon 2</td>
<td>2/18</td>
<td>0/96</td>
<td>rs7918614</td>
<td>Yes (6.7)</td>
<td>Yes (6.4)</td>
<td>Polymorphism</td>
</tr>
<tr>
<td>c.260-156T &gt; C</td>
<td>NA</td>
<td>Int 2</td>
<td>1/18</td>
<td>1/96</td>
<td>rs2839663</td>
<td>Yes (1.0)</td>
<td>No</td>
<td>Polymorphism</td>
</tr>
<tr>
<td>c.1578_1579insG</td>
<td>p.Val526X2</td>
<td>Exon 7</td>
<td></td>
<td></td>
<td>NA</td>
<td>No</td>
<td>No</td>
<td>Pathogenic mutation</td>
</tr>
<tr>
<td>c.2061A &gt; C</td>
<td>p.Pro687Pro</td>
<td>Exon 7</td>
<td>2/18</td>
<td>0/96</td>
<td>rs3486414</td>
<td>Yes (1.9)</td>
<td>Yes (1.94)</td>
<td>Polymorphism</td>
</tr>
<tr>
<td>c.2249C &gt; A</td>
<td>p.Ser750X</td>
<td>Exon 7</td>
<td></td>
<td></td>
<td>NA</td>
<td>No</td>
<td>No</td>
<td>Pathogenic mutation</td>
</tr>
</tbody>
</table>

---

In patients with keratoconus, we identified 7 heterozygous sequence variants; 3 nonsynonymous and 4 synonymous (Table 2). Three of these were known SNPs (rs80194531, rs7918614, and rs55238902) and one was a novel SNP (c.5177A > T; p.Pro1059Pro) seen in 5% (5/96) of controls, although absent from the 1KG and EVS datasets. There were 3 novel heterozygous sequence variants (c.1574G > A; p.Gly525-Glu, c.1920G > T; p.Gln640His, and c.2673G > C; p.Pro891-Pro), which were absent from controls (192 chromosomes), and the 1KG and ESP datasets.

The novel heterozygous sequence variant exon 7, c.1574G > A (p.Gly525Glu), was identified in a patient with sporadic keratoconus. This sequence variant results in a nonconservative substitution of a nonpolar neutral amino acid (glycine) with a polar negatively charged hydrophilic residue (glutamic acid). The glycine at position 525 is well conserved in primates, although in the chicken and shrew the residue is a glutamic acid. PMut predicted that the variant was pathologic with a score of 0.8 and high reliability, whereas PolyPhen-2 and SIFT predicted this substitution was benign and tolerated, respectively. Given this evidence, we proposed that p.Gly525-Glu is classified as potentially pathogenic. As the novel synonymous change c.2673G > C (p.Pro891Pro) was absent from controls, and the 1KG and ESP datasets, we sought to determine whether there was an impact on splicing due to the impact of the nucleotide substitution on exonic splicing enhancers (ESE) and silencers (ESS).31 On ESE analysis the nucleotide change c.2673G > C alters the predicted ESE binding site, but we were unable to obtain RNA from the patient to assess whether this nucleotide change results in aberrant ZEB1 splicing.

The novel heterozygous pathogenic mutation in exon 7, c.1920G > T (p.Gln640His), was identified in two siblings with keratoconus (Fig. 2B). The nucleotide change causes the conserved polar amino acid glutamine to be substituted by the positively charged, basic amino acid histidine. The glutamine at position 640 is the last amino acid of the homeodomain, and is well conserved across primates and other vertebrates as demonstrated in Figure 2C. SIFT analysis predicted that the variant was damaging, whereas PolyPhen-2 and PMut predicted this substitution was benign and neutral, respectively.
Figure 1. (A) Segregation analysis of the c.2249C > A, p.Ser750X ZEB1 mutation in a UK family affected by PPCD. (B) Slit-lamp examination of individual II:2 showed corneal endothelial vesicles seen typically in PPCD. (C) Sequence chromatogram showing the heterozygous c.2249C > A, p.Ser750X mutation in exon 7 of ZEB1 identified in the family affected by PPCD. FP, forward primer; RP, reverse primer. (D) Pentacam corneal topography images of right (OD) and left (OS) eyes of the proband (individual II:2). The upper maps show corneal steepening with regular astigmatism, but no signs of keratoconus. The lower maps show a general increase in corneal thickness (central corneal thickness 651 μm associated with PPCD).
TABLE 2. ZEB1 Sequence Variants Identified in the Keratoconus Patient Cohort

<table>
<thead>
<tr>
<th>Nucleotide Position</th>
<th>Amino Acid Change</th>
<th>Exon/Intron</th>
<th>Cases</th>
<th>Controls</th>
<th>dbSNP rs#</th>
<th>1KG (MAF %)</th>
<th>EVS (MAF %)</th>
<th>Classification</th>
</tr>
</thead>
<tbody>
<tr>
<td>c.192C&gt;T</td>
<td>p.Asp64Asp</td>
<td>Exon 2</td>
<td>1/70</td>
<td>0/96</td>
<td>rs7918614</td>
<td>Yes (6.7)</td>
<td>Yes (6.45)</td>
<td>Polymorphism</td>
</tr>
<tr>
<td>c.233A&gt;C</td>
<td>p.Asn78Thr</td>
<td>Exon 2</td>
<td>2/70*</td>
<td>0/96</td>
<td>rs80194531</td>
<td>Yes (1.2)</td>
<td>Yes (1.68)</td>
<td>Polymorphism</td>
</tr>
<tr>
<td>c.1257G&gt;A</td>
<td>p.Ala419Ala</td>
<td>Exon 7</td>
<td>1/70</td>
<td>0/96</td>
<td>rs5528902</td>
<td>Yes (1.2)</td>
<td>Yes (1.95)</td>
<td>Polymorphism</td>
</tr>
<tr>
<td>c.1574G&gt;A</td>
<td>p.Gly525Glu</td>
<td>Exon 7</td>
<td>1/70</td>
<td>0/96</td>
<td>NA</td>
<td>No</td>
<td>No</td>
<td>Potentially pathogenic</td>
</tr>
<tr>
<td>c.1920G&gt;T</td>
<td>p.Gln640His</td>
<td>Exon 7</td>
<td>1/70</td>
<td>0/96</td>
<td>NA</td>
<td>No</td>
<td>No</td>
<td>Pathogenic mutation</td>
</tr>
<tr>
<td>c.2673G&gt;C</td>
<td>p.Pro891Pro</td>
<td>Exon 8</td>
<td>1/70</td>
<td>0/96</td>
<td>NA</td>
<td>No</td>
<td>No</td>
<td>Potentially pathogenic</td>
</tr>
<tr>
<td>c.3177A&gt;T</td>
<td>p.Pro1059Pro</td>
<td>Exon 9</td>
<td>3/70</td>
<td>5/96</td>
<td>NA</td>
<td>No</td>
<td>No</td>
<td>Polymorphism</td>
</tr>
</tbody>
</table>

* Patients were African American and sequencing of ZEB1 in ethnically matched controls detected this variant in 3/23 controls.

Figure 2. (A) Segregation analysis of the c.1920G>T, p.Gln640His ZEB1 mutation in the family affected by keratoconus (siblings) and Fuchs' endothelial corneal dystrophy (mother). (B) Sequence chromatogram showing the heterozygous c.1920G>T, p.Gln640His mutation in exon 7 of ZEB1. (C) Multiple sequence alignment showing highly conserved glutamine at position 640 in exon 7 of ZEB1.
A. Individual II:1

B. Individual I:2

Figure 3. (A) Pentacam corneal topography images of right (OD) and left (OS) eyes of the proband (individual II:1; Fig. 2). The upper maps show inferior corneal steepening with a maximal corneal power of 56.6 D (OD) and 59.1 D (OS) associated with corneal thinning (bottom maps) confirming keratoconus. (B) Pentacam corneal topography corneal thickness maps of the mother of the proband (individual I:2; Fig. 2). The maps show a general increase in corneal thickness especially in the right eye (OD) associated with corneal endothelial pump failure and corneal hydration associated with FECD.
ZEB1 Gene in Corneal Dystrophies

Table 3. RT-qPCR of the Relative Expression Level of ZEB1 Transcriptional Target Genes in Cultured Corneal Keratocytes Harboring the ZEB1 Mutation p.Gln640His

<table>
<thead>
<tr>
<th>Gene</th>
<th>PPIA</th>
<th>HPRT1</th>
<th>RBL5</th>
<th>UBC</th>
<th>GAPDH</th>
<th>Average</th>
</tr>
</thead>
<tbody>
<tr>
<td>COL4A1</td>
<td>0.01</td>
<td>0.01</td>
<td>0.02</td>
<td>0.02</td>
<td>0.02</td>
<td>0.01</td>
</tr>
<tr>
<td>COL4A2</td>
<td>0.04</td>
<td>0.04</td>
<td>0.04</td>
<td>0.04</td>
<td>0.04</td>
<td>0.04</td>
</tr>
<tr>
<td>COL4A3</td>
<td>0.17</td>
<td>0.09</td>
<td>0.2</td>
<td>0.2</td>
<td>0.2</td>
<td>0.17</td>
</tr>
<tr>
<td>COL4A4</td>
<td>0.2</td>
<td>0.1</td>
<td>0.3</td>
<td>0.3</td>
<td>0.3</td>
<td>0.24</td>
</tr>
<tr>
<td>COL4A5</td>
<td>1</td>
<td>0.5</td>
<td>1.3</td>
<td>1.3</td>
<td>1.3</td>
<td>1.4</td>
</tr>
<tr>
<td>COL4A6</td>
<td>0.8</td>
<td>0.4</td>
<td>0.9</td>
<td>1.2</td>
<td>1.2</td>
<td>0.86</td>
</tr>
<tr>
<td>COL8A2</td>
<td>0.5</td>
<td>0.2</td>
<td>0.5</td>
<td>0.3</td>
<td>0.7</td>
<td>0.44</td>
</tr>
<tr>
<td>ZEB1</td>
<td>0.9</td>
<td>0.7</td>
<td>1.4</td>
<td>0.8</td>
<td>1.5</td>
<td>1.06</td>
</tr>
</tbody>
</table>

All mRNA levels were measured at CT threshold levels and were normalized with five reference genes: PPIA, HPRT1, RBL5, UBC, and GAPDH CT values. Values were expressed as fold increase over the corresponding values for control by the 2^ΔΔCT method.

Clinical Phenotype of Family With p.Gln640His ZEB1 Mutation

The proband harboring the p.Gln640His mutation was a 39-year-old Caucasian male (individual II:1, Fig. 2A) with bilateral keratoconus. He was diagnosed at age 25 and commenced contact lens wear at age 30. His best corrected visual acuity was of 20/40 (right eye) and 20/125 (left eye) with contact lenses. On slit-lamp biomicroscopy there was definite keratoconus; there were no other anterior segment abnormalities and the endothelium was normal. The minimal corneal thickness over the apex of the cone was 488 μm (right eye) and 448 μm (left eye). Keratometry measurements were 49.3/46.6 D (right eye) and 51.7/49.8 D in the left eye. Corneal topography confirmed keratoconus (Fig. 3A). The patient underwent a deep anterior lamellar keratoplasty in the left eye at the age of 40 years. The patient had a 37-year-old sister affected with bilateral keratoconus (individual II:2, Fig. 2A) who carried the same mutation. She was diagnosed with keratoconus at age 19 and underwent bilateral corneal transplantation at age 28 (right eye) and 31 (left eye). Their father had normal corrected visual acuities and corneal topography. Their mother was diagnosed with Fuchs’ endothelial corneal dystrophy (FECD) at age 56. FECD was diagnosed based on the presence of more than 5 mm confluent central corneal endothelial guttae in each eye (Krachmer grade 4 or higher). She underwent bilateral cataract surgery at age 58. At age 65, the patient had bilateral mild corneal epithelial and stromal edema associated with increased corneal thickness (Fig. 3B); central corneal thickness was 642 μm (right eye) and 618 μm (left eye). Her best corrected visual acuity was 20/40 (right eye) and 20/125 (left eye). There was no evidence of keratoconus clinically or on corneal topography. The patient underwent bilateral Descemet’s stripping automated endothelial keratoplasty at age 67. Segregation analysis identified the variant in both affected (keratoconus) siblings and the affected (FECD) mother, while it was absent in the unaffected father as shown in Figure 2A.

RT-qPCR of Mutant ZEB1 (p.Gln640His) Cultured Corneal Keratocytes

The expression of COL8A2 and the basement collagen genes (COL4A1, COL4A2, COL4A3, COL4A4, COL4A5, and COL4A6) was evaluated by RT-qPCR in cultured corneal keratocytes obtained from a patient harboring a missense mutation in ZEB1 (c.1920G>T, p.Gln640His, Table 3). COL4A1 and COL4A2 were markedly downregulated, and COL4A3, COL4A4, and COL8A2 were moderately downregulated. The expression level of ZEB1 was unchanged in the corneal keratocytes harboring the ZEB1 mutation.

Discussion

ZEB1 encodes the two-handed zinc-finger homeodomain transcription factor, which can act as either a transcriptional enhancer or repressor.32 Mutations in ZEB1 have been reported in posterior polymorphous corneal dystrophy (PPCD3; MIM #609141) and Fuchs’ endothelial corneal dystrophy (FECD6; MIM #613270), and herein we report a missense mutation in ZEB1 associated with keratoconus and highlight a genotype–phenotype correlation. Missense substitutions in the ZEB1 protein are associated with FECD6 and keratoconus, whereas protein truncating mutations result in PPCD3 (Fig. 4, Supplementary Table S3). Genotype-phenotype correlations also exist in TGFBI (MIM *601692) associated corneal dystrophies where specific missense mutations associated with arginine at codon 124 and 555 result in different phenotypic manifestations.33,34 The largest published series reported ZEB1 mutations in 25% of patients with PPCD (8/32 probands)13 and the prevalence of mutations in ZEB1 in PPCD has varied from 9.1% to 45.4%.5,9–11 In our study ZEB1 mutations were detected in 11.1% (2/18 probands) with PPCD. The majority of mutations were reported in exon 7, which most likely represents the relative size of this exon consisting of 1811 bps or 54% of the total ZEB1 transcript (3575 bps). All reported ZEB1 mutations have been unique, although the methionine at codon 1 mutation results from two different nucleotide substitutions; c.1A>G (p.Met1Val)10 and c.2T>G (p.Met1Arg).11 Although p.Met1Val and p.Met1Arg mutations are missense, it is predicted that they would result in a truncation of the ZEB1 protein as the initiation of translation codon is disrupted.10 All ZEB1 mutations reported in PPCD3 are nonsense, frameshift, and missense, which result in protein truncation.5,6,9–11 The proposed pathogenic mechanisms are haploinsufficiency either due to nonsense mediated decay or...
loss of functional domains due to truncations in the ZEB1 protein, or dominant-negative effects from the truncated mutant ZEB1 protein.5

Liskova et al. failed to detect an association of keratocan and ZEB1 mutations in their PPCD3 cohort.4 Three PPCD5 patients harbored keratocan dystrophy readings representing corneal steepening and increased corneal refractive power (corneal power > 49 D), and no corneal thinning or other signs of keratoconus.4 In our study, the keratoconus patients harboring a pathogenic missense ZEB1 mutation did not have evidence of PPCD3 and had characteristic features of keratoconus, including typical corneal topography. Missense mutations in ZEB1 have been reported in Fuchs’ endothelial dystrophy (FECD).14,15 although only identified in 2% (7/384) of cases in the largest published series.14 Genetic variation in the transcription factor 4 gene (TCF4; MIM602272) has been associated with the development of FECD, in a potential mechanism involving altered ZEB1 expression.35

We detected a pathogenic missense mutation in ZEB1 associated with keratocan and FECD segregating in a family suggesting common pathophysiological mechanisms. The dysregulation of type IV collagen represents a common link between ZEB1 mutation and the clinical phenotypes (PPCD3, FECD, and keratoconus). Mutations in type IV collagens (COL4A3, COL4A4, and COL4A5) cause Alport syndrome, a triad of nephropathy, sensorineural deafness, and eye features that include anterior lenticonus, PPCD, and keratoconus.36 Ectopic COL4A3 protein has been demonstrated in the corneal endothelium of an individual with PPCD3 resulting from a truncating mutation in ZEB1, and the expression of COL4A3 is increased in the corneal endothelium of ZEB1-null mice.37 The COL4A3 promoter contains a complex promoter binding site,37 and there is functional evidence that ZEB1 interacts with these proximal and distal E2-box motifs to regulate COL4A3 expression.38 Altered ZEB1-mediated COL4A3 transcriptional repression resulting from truncating mutations is proposed to result in PPCD3.38 In the corneal endothelium of a patient with PPCD3 resulting from a ZEB1 truncating mutation, qPCR demonstrated ZEB1 expression was reduced whereas COL4A3 expression was 4- to 5-fold higher than in the normal corneal endothelium.38 However, the actual site of the truncating ZEB1 mutation and the impact on ZEB1 protein structure was not reported.38 In our study, a missense mutation (p.Gln640His) in the homeodomain of ZEB1 resulted in a downregulation in type IV collagens, whereas the expression of ZEB1 mRNA was unaltered.

ZEB1 core and secondary binding sites have been reported in the promoter sites of COL8A2 and the basement collagen genes (COL4A1, COL4A2, COL4A3, COL4A5, and COL4A6), with only the COL4A3 promoter containing a complex site consisting of the core and proximal downstream secondary sequence.7 Given these findings, and that alterations in type IV collagen have been implicated in PPCD5,37,38 and corneal disease,36,39,40 we assessed the gene expression of COL8A2 and the basement collagen genes (COL4A1, COL4A2, COL4A3, COL4A4, COL4A5, and COL4A6) by RT-qPCR in cultured corneal keratocytes obtained from a patient harboring a missense mutation in ZEB1 (c.1920G > T; p.Gln640His). COL4A1, COL4A2, and COL4A3 expression were markedly reduced in corneal keratocytes harboring a missense ZEB1 mutation (p.Gln640His).

Reduced expression of COL4A1 and COL4A3 has been reported in keratocan.39,40 Sequencing COL4A3 and COL4A4 in 104 unrelated Slovenian patients with keratocan did not detect mutations, although there were significant differences in the genotypes of seven previously reported polymorphisms in COL4A3 and COL4A4 between keratocan patients and controls.35 COL4A1 mutations result in a variety of ocular defects, including abnormalities in the cornea and anterior segment.12,41 Mutational screening of COL4A1 in 15 Ecuadorian keratoconus families failed to detect pathogenic variants.44 Given the significant disruption in COL4A1 expression detected in our study and previously in keratocan,40 further work is required to determine the role of COL4A1 in keratoconus. We detected a reduction in COL8A2 expression in corneal keratocytes harboring a missense ZEB1 mutation (p.Gln640His). Disruption of COL8A1 and COL8A2 in a knockout mouse model resulted in corneal thinning: keratoglobus.45 Missense mutations in COL8A2 result in PPCD and FECD8, although no pathologic variants have been reported in keratoconus.46

The findings reported herein and in the literature support the proposal that missense substitutions in the ZEB1 protein are associated with FECD6 and keratoconus, whereas protein truncating mutations result in PPCD3. We propose that the functional impact of the class of mutation on ZEB1 alters downstream gene expression and this forms the basis of the genotype–phenotype correlation. The identification and characterization of the molecular pathways regulated by ZEB1 in the cornea and the functional impact of mutations on these pathways is required to understand fully the observed genotype–phenotype correlations. Keratoconus and FECD result in significant ocular morbidity and are major indications for corneal transplantation.47 Dissecting the role of ZEB1 in the pathogenesis of inherited corneal dystrophies will provide important insights into maintenance of corneal shape and clarity.

Acknowledgments

The authors thank the patients who participated in our study, as well as Jude Lynch, who provided technical assistance with the construction of illustrations; Khanh-Nhat Tran-Viet (Duke University Medical Center) for assistance with DNA shipping; and Gail Billingsley and Ajoy Vincent (The Hospital for Sick Children, Toronto) for collecting clinical information.

Supported by the Research and Development Office (Grant RRG Grant 4.46), Northern Ireland, Department of Education and Learning (DEL), Northern Ireland, and Fight for Sight (UK). JL is a Fight for Sight PhD student.

Disclosure: J. Lechner; None; D.P. Dash; None; D. Muszynska; None; M. Hossceni; None; F. Segev; None; S. George; None; D.G. Frazer; None; J.E. Moore; None; S.B. Kaye; None; T. Young; None; D.A. Simpson; None; A.J. Churchill; None; E. Héon; None; C.E. Willoughby; None

References

6. Nguyen DQ, Hossceni M, Billingsley G, Heon E, Churchill AJ. Clinical phenotype of posterior polymorphous corneal dys-


