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Smac-Derived Aza-Peptide As an Aminopeptidase-Resistant XIAP BIR3 Antagonist

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Abstract: The peptidic nature of anti-IAPs N-terminus Smac-derived peptides precludes their utilization as potential therapeutic anticancer agents. Recent advances in the development of novel Smac-derived peptidomimetics and non-peptidic molecules with improved anti-IAPs activity and resistance to proteolytic cleavage have been reported and led to a number of candidates that are currently in clinical trials including LCL-161, SM-406/AT-406, GDC-0512/GDC-0917, and birinapant. As an attempt to improve the proteolytic stability of Smac peptides, we developed the Aza-peptide AzaAla-Val-Pro-Phe-Tyr-NH2 (1). Unlike unmodified peptide Ala-Val-Pro-Phe-Tyr-NH2 (1), analogue (2) exhibited resistance towards proteolytic cleavage by two aminopeptidases; LAP and DPP-IV, while retaining its IAP inhibitory activity. This was due to the altered planar geometry of the P1 residue side chain. Our findings showed that using azas-isosteres of bioactive peptide sequences imbue the residue with imperviousness to proteolysis; underscoring a potential approach for developing a new generation of Smac-derived Aza-peptidomimetics.

Keywords: Aminopeptidases, Apoptosis, Aza-peptides, Caspase 9, IAPs, Smac.

INTRODUCTION

Inhibition of the anti-apoptotic IAPs proteins by short peptides derived from the N-terminal sequence of the pro-apoptotic mitochondrial protein Smac is an emerging strategy for re-engaging apoptosis in resistant cancer cells over-expressing IAPs. Although Smac-derived peptides have been demonstrated to augment the activity of anticancer drugs both in vitro and in vivo [1, 2], their entirely peptidic nature almost certainly precludes their utilization as anticancer therapeutic agents. This is because of the innate and generally impaired bioavailability of peptide ligands mainly due to their instability towards proteolytic cleavage. There have been long standing efforts in developing protease-resistant Smac-derived peptidomimetics (monovalent and bivalent) and non-peptidic analogues by ourselves [3] and many other groups [4-9] that led to in some cases to IAP inhibitors that reached clinical trials [10] such as LCL-161, SM-406/AT-406, GDC-0512/GDC-0917, and birinapant. As an attempt to improve the proteolytic stability of Smac peptides, we developed the Aza-peptide AzaAla-Val-Pro-Phe-Tyr-NH2 (1). Unlike unmodified peptide Ala-Val-Pro-Phe-Tyr-NH2 (1), analogue (2) exhibited resistance towards proteolytic cleavage by two aminopeptidases; LAP and DPP-IV, while retaining its IAP inhibitory activity. This was due to the altered planar geometry of the P1 residue side chain. Our findings showed that using azas-isosteres of bioactive peptide sequences imbue the residue with imperviousness to proteolysis; underscoring a potential approach for developing a new generation of Smac-derived Aza-peptidomimetics.

Keywords: Aminopeptidases, Apoptosis, Aza-peptides, Caspase 9, IAPs, Smac.
modified peptide AzaAla-Val-Pro-Tyr-NH₂ (2) could still make the important electrostatic contacts within the XIAP BIR3 binding pocket, by virtue of the retained and unmodified primary amino function of the AzaAla residue, whilst exhibiting enhanced stability towards aminopeptidases (exemplified mainly by LAP). All this is due to the altered geometry at P1, where the side chain residue of the SP₂ hybridised L- amino acid (Fig. 2). This letter thus reports on the synthesis of the Smac-derived Aza-peptide (2), testing its stability towards LAP compared to peptide (1), and an examination of its ability to re-engage caspase-9 activation for apoptosis in cell lysates obtained from MDA-MB-231 breast cancer cells.

![Figure 2. Configuration of L-Ala and AzaAla residues.](image)

**METHODS**

**General Protocol for the Microwave Assisted Solid Phase Peptide Synthesis (MW-SPPS)**

Peptides were synthesized using MW-SPPS on Rink Amide MBHA resin (substitution 0.65 mmol/g, 100-200 mesh) and were purified by RP-HPLC as previously described [3].

To a stirred solution of 1-(tert-butoxycarbonyl)-2-(9H-fluoren-9-methoxycarbonylamino)-1-methylhydrazine (4) in THF/H₂O (70:30 mL) was added solid sodium bicarbonate (9.3 g, 110.7 mmol) followed by a solution of Fmoc-OSu (20.3 g, 60.76 mmol) in THF (140 mL). The mixture was stirred overnight at room temperature, after which the THF was separated and the aqueous phase was washed with diethyl ether (3 x 25 mL). The combined organic extracts were washed with H₂O, separated, dried over anhydrous magnesium sulphate, filtered and concentrated under reduced pressure to afford yellow viscous oil (18 g) that was directly used in the next step without purification.

1-(tert-Butoxycarbonyl)-2-(9H-fluoren-9-methoxycarbonylamino)-1-methylhydrazine (4)

TFA (50 mL) was added to the yellow oil (4) obtained above and the reaction mixture was stirred, at room temperature, overnight. The reaction mixture was then concentrated under vacuum in a well-ventilated fume hood to give reddish viscous oil. The residual oil was dissolved in H₂O/ethyl acetate mixture (50/30 mL), stirred at room temperature and sodium bicarbonate solid was added till no effervescence was observed (pH 9). A white precipitate was formed, which was filtered, washed with ethyl acetate and dried under vacuum to afford 12.56 g (84.6%) of 2-(9H-Fluoren-9-methoxycarbonylamino)-1-methylhydrazine (5) as a white powder of melting point 145-150°C. ¹H NMR (CDCl₃, 400 MHz) δ 7.94-7.32 (m, 8H), 6.24 (br s, 1H), 4.40 (d, 2H), 4.16 (t, 1H), 2.58 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 157.13, 143.73, 141.36, 127.76, 127.07, 124.97, 120.01, 67.01, 47.24, 39.29; ESI-Mass m/z 291.3 [M + Na⁺].

Coupling of 1-chlorocarbonyl-2-(9H-Fluoren-9-methoxycarbonylamino)-1-methylhydrazine (6) to Val-Pro-Phe-Tyr-Resin (7) (Scheme 1, Route A)

To a 50 mL round bottom flask, containing a cold solution (-10°C) of triphosgene (0.66 mmol, 196 mg) and 2-(9H-Fluoren-9-methoxycarbonylamino)-1-methylhydrazine (5) (2 mmol, 536 mg) in 15 mL THF, was added drop wise N-methylmorpholinium chloride was formed. The reaction mixture was centrifuged at 2000 x g for 5 minutes and the supernatant was separated and concentrated under vacuum. 1-chlorocarbonyl-2-(9H-Fluoren-9-methoxycarbonylamino)-1-methylhydrazine (6) residue 500 mg (1.5 mmol, 75% yield) was obtained, ESI-Mass for sample dissolved in methanol m/z 328.5 [Moc-NH-Ni(CH₃)₂-COOC(CH₃)₃]. The residue was then used for the microwave-assisted coupling to H₃N-Val-Pro-Phe-Tyr-Resin (7) with DIEA as activator base, where 1-chlorocarbonyl-2-Fmoc-1-methylhydrazine (DEA 1/2). The reaction was repeated for 7 cycles (coupling completion was monitored by Kaiser Test); each coupling cycle was performed for 10 minutes, at 22 Watts, at a temperature of 75°C. After coupling completion, the peptide was cleaved from the resin and puri-
fied using RP-HPLC (check under general protocols for the MW-SSPs) to obtain AzaAla-Val-Pro-Phe-Tyr-NH₂ (2) (8% yield and 98% purity after semi-preparative RP-HPLC purification). ESI-Mass m/z: 618.6 [M+Na⁺].

Coupling of 2-(9H-Fluoren-9-methoxycarbonylamo)-1-methylhydrazine (5) to Val-Pro-Phe-Tyr-Resin (7) (Scheme 1, Routes B and C)

For 0.25 mM scale microwave-assisted synthesis of AzaAla-Val-Pro-Phe-Tyr-NH₂ (2), 2-(9H-Fluoren-9-methoxycarbonylamo)-1-methylhydrazine (5) (15 mM, 402 mg, 6 equivalent) and carbonyldimidazol (1.5 mM, 243 mg, 6 equivalent) or p-nitrophenyl chloroformate (1.5 mM, 302 mg, 6 equivalent) were dissolved in DMSO (6 mL). DIEA was used as activator base, where 2-Flmoc-1-methylhydrazine (5) (DEA carbonyldimidazol or p-nitrophenyl chloroformate (11/2) for one coupling cycle. The coupling reagents were transferred to the microwave reaction vessel for coupling (5) to the resin bound peptide H₂N-Val-Pro-Phe-Tyr-Resin (7). The reaction was repeated for 7 cycles (coupling completion was monitored by Kaiser Test and ESI-MS); each coupling cycle was performed for 10 minutes, at 22 Watts, at a temperature of 75 °C.

Aminopeptidase Susceptibility Assays

In 1.5 mL microcentrifuge tube, 940 µL of assay buffer (DPP-IV assay buffer: 25 mM Tris, 140 mM NaCl and 10 mM KCl at pH 8.5; LAP assay buffer: 200 mM NaCl at pH 8.5) and 10 µL of peptide DMSO solution (5 mM final concentration) were added per well. A FLUOstar OPTIMA spectrofluorimeter (BMG LABTECH, Ortenberg, Germany) was used for excitation wavelength of 380-400 nm and emission wavelength of 470-500 nm to determine the caspase-9 activity through the Ac-LEHD-AFC substrate cleavage with the release of 7-amino-4-trifluoromethylcoumarin fluorescence moiety. The reaction was monitored for 1 h. Reactions were carried out in triplicates for accuracy. Finally, percent caspase-9 release was calculated from the following formula:

% caspase 9 derepression = 100 X (slope of treatment curve / slope of maximum activation (dTAP + cyt.c) curve fit)

RESULTS AND DISCUSSION

Synthesis of AzaAla-Val-Pro-Phe-Tyr-NH₂ (2)

The Microwave-assisted solid phase synthesis (MW-SSPS) of the target peptide included the coupling of 2-(9H-Fluoren-9-methoxycarbonylamo)-1-methylhydrazine (5) to the tetrapeptide sequence Val-Pro-Phe-Tyr-tethered to the resin (7) (Scheme 1), but first the synthesis of (5) was required. Compound (5) was synthesized from 1-(tert-butoxycarbonyl)-1-methylhydrazine (3), using the previously reported synthesis protocol by Busnel et al. [22] and others [23-25]. The synthesis comprises two steps, an initial Fmoc-protection of the primary amino function of the hydrazine, followed by Boc removal of the secondary amine functionality (Scheme 1). The introduction of the Fmoc-group was accomplished using Fmoc-OBu, in the presence of sodium bis carbonate, to obtain (4), which was then treated with TFA in order to effect the removal of the Boc-group. The final product, N-Fmoc-methylhydrazine (5), was obtained in high yield (85%, based on the starting amount of 4).

The synthesis of the target Aza-peptide (2) was achieved as shown in Route A (Scheme 1) through the activation of (5) by treatment with triphosgene in the presence of N₃₅ methyl morpholine, at -10 °C, in THF. The product, 1-chloro-carbonyl-2-(9H-Fluoren-9-methoxycarbonylamo)-1-methylhydrazine (6) was obtained in high yield (75% based on the amount of starting material 5) and was used directly to acylate the resin-bound H₂N-Val-Pro-Phe-Tyr-sequence (7) using microwave-assisted coupling, employing DIEA as activator base (Scheme 1, Route A). Samples of resin were subjected to the Kaiser Test and ESI-MS, in order to assess the extent of coupling (6). It was found that seven repeat coupling cycles (10 minutes/cycle, 22 Watts, 75 °C) were required for completion of the reaction.

Boeglin and Lubell have previously shown that coupling of various Fmoc-aza-amino acid chlorides (including the N-terminus amino group of resin tethered peptides) is only complete after 6 h reaction in DCM with DIEA as activator base using conventional SPPS, and in some cases the coupling has to be repeated to ensure completion [24]. Employing microwave-assisted coupling, we managed to significantly reduce the coupling reaction time to 70 minutes (7 cycles x 10 minutes/cycle). Unfortunately, even with this repetitive coupling protocol, (2) could only be obtained as a low purity (~25%) crude product, as estimated by analytical
RP-HPLC (data not shown). This could be due to the infeasibility of purifying the highly active and unstable intermediate (8), which necessitates using it as a crude product for coupling to the resin bound peptide (7). However, the crude Aza-peptide (2) was purified successfully, by semi-preparative RP-HPLC, to give a final product with a yield of only 8%, but of 98% purity (Fig. 3).

Triphosgene is not compatible for use within a microwave-assisted synthesizer, due to it being highly reactive and with potential for interaction with the amide groups of the microwave compatible solvents DMF and NMP. Therefore in attempts to improve the yield and purity of the product and to automate the synthesis, additional carbonylating agents were tried for coupling N-Fmoc-methylhydrazine (5).
to the N-terminal amino group of the resin-tethered peptide (7), under microwave conditions. We tried both para-nitrophenyl chloroformate (Scheme 1, Route B) and carbonyldiimidazole (Scheme 1, Route C) mediated carbonylation and coupling using the conditions described in the experimental section. Disappointingly, we did not obtain any of the target peptide, even after 7-repeat coupling cycles in case of para-nitrophenyl chloroformate (data not shown) and incomplete coupling was observed when carbonyldiimidazole was used as activator, as indicated by ESI-MS analysis (Fig. 4). Similar observations have been reported by Han et al., in the attempted incorporation of two aza-amino acids into target sequences when using para-nitrophenyl chloroformate and carbonyldiimidazole [26]. This could be due to a low inherent reactivity of the respective aryl carbamates and N-carbonyl imidazoles, or the poor accessibility of these derivatives to the amino functionality of the tethered peptide within the resin matrix, as reported by André et al. [27]. Ultimately, the AzaAla-Val-Pro-Phe-Tyr-NH₂ (2) synthesis was only successful through the triphosgene-mediated carbonylation approach (Scheme 1, route A).

Aminopeptidase Susceptibility

Compound (2) was then tested for its stability against two aminopeptidases; namely LAP and DPP-IV, and in comparison to the wild-type Smac-derived native sequence Ala-Val-Pro-Phe-Tyr-NH₂ (1) and the N-methyl alanyl analogue NMe-Ala-Val-Pro-Phe-Tyr-NH₂ (9).
Figure 4. ESI-MS trace for crude AzaAla-Val-Pro-Phe-Tyr-NH₂ (2) produced by carbonyldiimidazole-mediated carbonylation of N-Fmoc-methylhydrazine (5) showing peaks of unreacted Val-Pro-Phe-Tyr-NH₂ even after 7-repeated coupling cycles, indicating incomplete coupling.

LAP is an aminopeptidase that can cleave N-terminal Ala residues from peptides [28]. As anticipated, the P1 Ala residue was completely hydrolysed from the native peptide sequence (1) in 30 minutes, to give the truncated hydrolysis product Val-Pro-Phe-Tyr-NH₂, as shown by RP-HPLC and ESI-MS (Fig. 5 (a)). Strikingly, the AzaAla-analogue (2) was completely resistant to hydrolysis after 6 h of exposure to LAP (Fig. 5 (b)), as was the case with the N-methyl alanyl protected analogue (9) (Fig. 5 (c)). These results suggest that designing Smac-derived analogues with a planar SP² hybridised Aza-alanyl modified N-terminal will improve the proteolytic resistance for those candidates, making them as resistant as the widely used N-methyl alanyl congeners.

Interestingly, the Smac-derived pentapeptide analogue (1), AzaAla-analogue (2) and N-Me-Ala-analogue (9) proved to be resistant to aminopeptidase DPP-IV after 6 h exposure to the enzyme, as observed by RP-HPLC and ESI-MS (data not shown). Although DPP-IV is an N-terminal dipeptidyl
peptidase that can cleave a dipeptide unit in ‘one bite’ from longer peptides with Val at the P2 position [29], it might have been expected that all the three Smac-derived peptides (1), (2) and (9) would have been subject to degradation by this peptidase (they all have Val at P2). However, the presence of Pro at P3 makes all of the N-terminal Smac-derived sequences intrinsically resistant to DPP-IV, since peptides with Pro at P3 are known to function as DPP-IV inhibitors [30].

IAP Antagonistic Activity

Subsequently, the antagonistic activity of AzaAla-analogue (2) against XIAP BIR3 was assessed by caspase-9 activity recovery assay. In this assay, the ability of the Smac-derived analogues to reactivate caspase-9 in the presence of its inhibitor rhXIAP BIR3 (500 nM) was determined by the increase in caspase-9 activity that they induced.

Remarkably, the Aza-peptide analogue (2) retained the ability to antagonise the inhibitory effect of XIAP BIR3 on caspase-9 activation as effective as the wild-type (1) and the N-Methyl-Ala-sequence (9) (Fig. 6). In essence, (2) was able to restore 36.2 ± 4.9% of caspase-9 activity in the presence of 500 nM rhXIAP BIR3, at 1 µM compared to 43.4 ± 5.2% for the wild sequence (1) and 51.4 ± 7.1% for the N-Me-Ala-analogue (9), at the same concentration (mean ± SD; n=3) (Fig. 6). These results demonstrate that the planar conformation of the SP2 hybridised AzaAla residue did not dramatically mislocate the side-chain methyl group at P1 permitting the essential hydrophobic interaction with the indole ring of the W310 residue of XIAP BIR3 (Fig. 1[3]). Moreover, analogue (2) can still form the essential electrostatic interaction with the proximal Glu314 of XIAP BIR3, owing to the retained and unmodified N-terminal primary amino function of the AzaAla residue (Fig. 1[3]).

CONCLUSION

In the light of the above results, the modified peptide AzaAla-Val-Pro-Phe-Tyr-NH2 (2) was a successful bio-isostere for the N-terminal Smac-derived pentapeptide parent sequence Ala-Val-Pro-Phe-Tyr-NH2 (1). The Aza-peptide (2) is an equipotent analogue for (1) as a XIAP BIR3 inhibitor, whilst exhibiting enhanced stability towards aminopeptidases, exemplified mainly by LAP, owing to the planar geometry of the Aza-Ala residue at P1. Although the reported Aza-peptide (2) is not a therapeutic candidate per se, due to lack of cell permeability, it represents an important lead compound for the development of a new class of anti-cancer Smac-derived Aza-peptidomimetic therapeutic candidates. Our current studies focus on developing libraries of these novel candidates, where the N-terminal Ala residue at P1 is occupied with the Aza-Ala isostere instead of the widely used N-methyl substituted Ala, with P2, 3 and 4 residues modified as previously reported by ourselves [3] and others [4]. These structural modifications of the asing Smac-derived Aza-peptidomimetics are expected to improve the efficacy and proteolytic stability of these therapeutic candidates.

LIST OF ABBREVIATIONS

IAPs = Inhibitor of Apoptosis Proteins
Smac = Second mitochondria-derived activator of caspases
XIAP BIR3 = X-linked inhibitor of apoptosis Protein-Baculovirus IAP Repeat-3 domain
SAR = Structure Activity Relationship
LAP = Leucine Aminopeptidase
DPP-IV = Dipeptidyl Peptidase-IV

CONFLICT OF INTEREST

The authors declare there is no conflict of interest for this research work.

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