Generation and Characterization of ALX-0171, a Potent Novel Therapeutic Nanobody for the Treatment of Respiratory Syncytial Virus Infection

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Respiratory syncytial virus (RSV) is an important causative agent of lower respiratory tract infections in infants and elderly individuals. Its fusion (F) protein is critical for virus infection. It is targeted by several investigational antivirals and by palivizumab, a humanized monoclonal antibody used prophylactically in infants considered at high risk of severe RSV disease. ALX-0171 is a trimeric Nanobody that binds the antigenic site II of RSV F protein with subnanomolar affinity. ALX-0171 demonstrated in vitro neutralization superior to that of palivizumab against prototypic RSV subtype A and B strains. Moreover, ALX-0171 completely blocked replication to below the limit of detection for 87% of the viruses tested, whereas palivizumab did so for 18% of the viruses tested at a fixed concentration. Importantly, ALX-0171 was highly effective in reducing both nasal and lung RSV titers when delivered prophylactically or therapeutically directly to the lungs of cotton rats. ALX-0171 represents a potent novel antiviral compound with significant potential to treat RSV-mediated disease.

Human respiratory syncytial virus (RSV) is the most important viral pathogen causing acute lower respiratory tract infections in infants worldwide and is estimated to result in ~3.4 million yearly hospitalizations and ~200,000 deaths globally (1). RSV typically causes its primary infection at the point of entry: apical ciliated epithelial cells that line the nasal cavity and airways (2, 3). Primary infections are usually asymptomatic, with clinical signs ranging from mild upper respiratory tract illness to severe lower respiratory tract infections, including pneumonia and bronchiolitis (4). In addition to the acute consequences of infection, the development of long-term recurrent wheezing and asthma has been associated with severe RSV infections in infancy (5, 6). Despite the major clinical importance of RSV, no vaccines or widely accepted antiviral therapies are currently available. The only available drug specific for human RSV is palivizumab (Synagis), a marketed monoclonal antibody that is administered prophylactically before and during the RSV season to infants at high risk of having severe human RSV disease (7–9). Its use is restricted to premature infants (gestational age, <29 weeks), if they have no other underlying morbidities, and infants with chronic lung disease, congenital heart disease, or a compromised immune system during the first year of life (10).

RSV is a member of the Pneumovirinae genus of the Paramyxoviridae family and has a linear single-stranded, nonsegmented RNA molecule of negative polarity as its genome. This genome contains 10 genes which encode 11 proteins. The transmembrane glycoproteins F and G are the primary surface antigens of RSV. The attachment (G) protein mediates binding to cell receptors, while the F protein promotes fusion of the viral and cell membranes, allowing virus entry into the host cell cytoplasm (11). The F protein also promotes the fusion of infected cells with adjacent uninfected cells, facilitating the formation of multinucleated cell formations (syncytia), which allow cell-to-cell transmission of the replicated viral RNA and confer additional protection for the virus against host immune responses (12).

On the basis of the antigenic and genetic variability of the G protein, two subgroups of RSV (subgroups A [RSV-A] and B [RSV-B]) have been identified, and these are composed of evolving genotypes (13–17). In contrast to the G protein, the F protein is mostly conserved between RSV subgroups A and B (89% amino acid identity) and is therefore considered the most promising target for the development of viral entry inhibitors.

Nanobodies are therapeutic proteins derived from the heavy-chain variable domains (VHH) that occur naturally in heavy chain-only immunoglobulins of the Camelidae (18, 19). The formatting flexibility of Nanobodies into multivalent constructs, their small size, their stability (which allows delivery through nebulization), and their ease of production make their use against viral targets appealing (20–22).

Received 31 July 2015 Returned for modification 19 August 2015 Accepted 25 September 2015 Accepted manuscript posted online 5 October 2015


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Supplemental material for this article may be found at http://dx.doi.org/10.1128/AAC.01802-15.

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Here we describe the in vitro and in vivo characterization of ALX-0171, a trivalent Nanobody composed of three monovalent Nb017 moieties linked together with glycine-serine (GS) linkers which is designed to target the RSV F protein for delivery via inhalation. ALX-0171 is currently in clinical development for the treatment of RSV infections in infants (23).

MATERIALS AND METHODS

Generation of RSV-specific Nanobodies. Monovalent RSV F protein-specific Nanobodies were identified from immune libraries of llamas that received repetitive injections with soluble recombinant F protein (F_2p,N:N protein), inactivated RSV-A (catalog number RSV79; HyTest), or a combination of both antigens. The soluble recombinant form of the F protein (derived from the Long strain) was produced with recombinant Sendai virus in embryonated eggs (24). RSV-neutralizing Nanobodies were identified by screening 162 distinct clones in an in vitro microneutralization assay with RSV subtype A Long on HEp-2 cells. RSV-neutralizing Nanobodies were formatted into multivalent constructs by genetic fusion with flexible Gly-Ser (GS) linkers of different lengths. Monovalent RSV Nanobody was routinely generated as C-terminal c-myc- His-tagged proteins in Escherichia coli (22). For the production of untagged monovalent and trivalent Nanobody constructs, a Pichia pastoris X-33 expression system was used (see Fig. S1 in the supplemental material). ALX-0171 is a trimeric Nanobody consisting of three RSV-targeting Nb017 subunits linked by two GS linkers.

Competition enzyme-linked immunosorbent assay (ELISA). Ninety-six-well microtiter plates (MaxiSorp immunoplates; Nunc) were coated overnight with inactivated RSV-A (HyTest) at 3.5 μg/ml in bicarbonate buffer. The plates were blocked for 1 h with SuperBlock T20 (phosphate-buffered saline [PBS]) blocking buffer (Pierce) before being washed 3 times with PBS–0.05% Tween 20. Dilutions of ALX-0171 or palivizumab were mixed with 10 nM biotinylated ALX-0171 (1:1 ratio) and immediately transferred to the plates. After overnight incubation at room temperature (RT) and appropriate washing, a 1/3,000 dilution of peroxidase-labeled streptavidin (Thermo Scientific) was added to the plates for 1 h at RT. Bound streptavidin-horseradish peroxidase was revealed by adding 100 μl/well of enhanced soluble tetramethylbenzidine (esTMB; SDT Reagents) for 10 min, followed by 1 N HCl. The absorbance was read at 450 nm, and the reference wavelength was 620 nm.

SPR. Affinity constants (the equilibrium dissociation constants [KD]) of Nb017, ALX-0171, and palivizumab were determined by surface plasmon resonance (SPR) analysis on a Biacore X100 instrument (GE Healthcare) was amine coupled to a CM5 sensor chip at a pH 4.5. Approximately 70 RU of His-tagged prefusion F protein was then bound to the anti-His monoclonal antibody. Compounds (ALX-0171, Nb017, Fab D25, Fab 2F, and palivizumab) were injected at 5 different concentrations (1 to 16 nM for ALX-0171, 8 to 128 nM for Nb017, 4 to 64 nM for palivizumab, 1.25 to 20 nM for Fab 2F and Fab D25) at a flow rate of 30 μl/min. The association and dissociation phases were 2 and 13 min, respectively (for Fab D25, the dissociation time was increased to 30 min). The chip was regenerated using 10 mM glycine, pH 1.5. Different concentrations of the compounds were used to generate binding curves, which were fitted to a 1:1 Langmuir binding model for the calculation of the kinetic parameters association rate (k_on) and dissociation rate (k_off). The K_D was then calculated as the ratio of these two rate constants (k_off/k_on).

Binding of Nanobodies to RSV escape mutant F protein. HEP-2 cells grown in 100-mm-diameter plates were infected with either wild-type RSV (the RSV Long strain) or previously described mutant viruses resistent to neutralization by monomolecular antibodies (11, 28, 29) (multiplicity of infection, 1 to 3 PFU/cell). After 48 h, the cells were harvested and extracts were made in 500 μl buffer/plate (the buffer was 10 mM Tris-HCl, pH 7.5, 150 mM NaCl, 5 mM EDTA, and 1% octyl-glucoside). Equal amounts of each extract (50 μl of a 1:500 dilution in PBS) were used to coat 96well microtiter plates overnight at 4°C. The wells were saturated with 1% bovine serum albumin in PBS for 1 h, and after washing with PBS, Nanobodies were added to the wells and the plates were incubated for 1 h at RT. After washing, bound Nanobody was detected with a 1:1,000 dilution of rabbit anti-llama IgG antibody, followed by a 1:200 dilution of peroxidase-conjugated donkey anti-rabbit polyclonal antibody. o-Phenylenediamine dihydrochloride (OPD; Sigma) was used as the substrate, and reactions were stopped with 3 N H2SO4 after the absorbance at 490 nm was measured.

RSV clinical strains. Original clinical isolates of RSV-A and RSV-B strains were selected from the RSV bank at the Baylor College of Medicine (Houston, TX). Three additional RSV-A isolates (BT2a, BT3a, BT4a) were isolated from infants hospitalized with bronchiolitis in the Royal Belfast Hospital for Sick Children. This study was approved by The Office for Research Ethics Committees Northern Ireland (ORECNI). Written informed parental consent was obtained. They were all isolated and cultured in HEP-2 cells, as previously described (30, 31).

Antiviral assays. The neutralization activity of the selected Nanobodies and palivizumab was measured in a microneutralization assay, a neutralization assay, and a plaque reduction assay. They were performed using HEP-2 cell monolayers, as described in the supplemental material.

Cotton rat studies. Male and female cotton rats were bred and housed in the Baylor College of Medicine’s vivarium in cages covered with barrier filters and given food and water ad libitum. Cotton rats weighing between 60 to 125 g were lightly anesthetized with isoflurane and intranasally challenged with 100 μl of 1.41 X 10^6 to 5.3 X 10^5 PFU of RSV Tracy, depending on the experiment. RSV Tracy stocks were grown in HEP-2 cells. All the experimental protocols were approved by the Baylor College of Medicine Investigational Animal Care and Use Committee.

ALX-0171 was administered either intranasally or by nebulization. For intranasal administration, cotton rats were lightly anesthetized with isoflurane and inoculated intranasally with 100 μl of ALX-0171. For nebulization, an Akita® Apixneb nebulizer (Vectora GmbH, Germany) was used for whole-body exposure (see Fig. S2 in the supplemental material).

Cotton rats were euthanized with CO2 at 4 days postinfection. The large left lobe and one of the large right lobes of the lungs were removed, rinsed in sterile water, and weighed. The left lobe was then transpleurally lavaged using 3 ml of Iscove’s medium with 15% glycerol mixed with 2% fetal bovine serum (FBS)–minimal essential medium (MEM) (1:1, vol/vol) in a 3-ml syringe with a 26-gauge by 3/8-in. needle and injected at multiple sites to totally inflate the lobe. Subsequently, the lavage fluid was recovered by gently pressing the inflated lobe flat and further used to transpleurally lavage the right lobe following the same technique. The lavage fluid was collected and stored on ice until titrated. To obtain nasal washes, the jaws were disarticulated, the head was removed, and 1 ml of Iscove’s medium with 15% glycerol mixed with 2% FBS–MEM (1:1, vol/vol) was pushed through each nare (total, 2 ml). The effluent was collected from the posterior opening of the pallet and stored on ice until titrated. RSV Tracy lung lavage titers and nasal wash titers were determined by plaque assay as described in the supplemental material.

Statistical analysis. A one-way analysis of variance (ANOVA) followed by a post hoc Dunnett’s test for pairwise comparison was performed to compare the treated groups and the buffer-treated groups. A P value of <0.05 was considered statistically significant. Statistical analysis was performed with GraphPad Prism (version 5) software.

RESULTS

Generation and production of ALX-0171. The development of a Pichia pastoris strain for the production of ALX-0171 delivered a stable recombinant strain suitable for the manufacture of the multivalent trimeric ALX-0171 Nanobody. Further streamlining of the typical Pichia pastoris fermentation process resulted in an
ALX-0171 upstream process in which more than 7.5 g/liter ALX-0171 was secreted in the fermentation broth. After clarification of the broth, the downstream process consisted of capture, intermediate purification, and polish chromatography steps and was followed by a final formulation step. A range of ALX-0171 concentrations was tested in combination with a series of buffers and excipients to provide the highest stability after storage, nebulization, freeze-thaw, etc. These formulation and (stressed) stability studies resulted in the formulation of ALX-0171 as a stable nebulizer solution.

**Characteristics of ALX-0171 binding to RSV F protein.** The kinetics of Nb017 and ALX-0171 binding to the RSV F protein in its prefusion conformation was assessed by SPR analysis. The conformational integrity of the F protein was confirmed with the antigenic site ß-specific Fab D25, which bound only to the prefusion conformation, whereas the antigenic site 1-specific Fab 2F did not bind to the prefusion conformation, as expected. The binding affinity of palivizumab was 0.88 nM on the prefusion conformation, whereas the antigenic site ß-specific Fab 2F did not bind to the prefusion conformation, as expected. The binding affinity of palivizumab was 0.88 nM on the prefusion conformation, whereas the antigenic site ß-specific Fab 2F did not bind to the prefusion conformation, as expected. The binding affinity of palivizumab was 0.88 nM on the prefusion conformation, whereas the antigenic site ß-specific Fab 2F did not bind to the prefusion conformation, as expected. 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ALX-0171, a Novel Nanobody Targeting RSV

Table 2: Neutralization of RSV prototypic and clinical isolates

<table>
<thead>
<tr>
<th>RSV strain</th>
<th>IC_{50} Mean ± SD IC_{50} (nM)</th>
<th>VN_{so} Mean ± SD VN_{50} (nM)</th>
<th>Fold difference</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>ALX-0171</td>
<td>Palivizumab</td>
<td>Palivizumab</td>
</tr>
<tr>
<td>RSV-A</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Long</td>
<td>0.1 ± 0.07 (n = 3)</td>
<td>12.6 ± 5.4 (n = 3)</td>
<td>126</td>
</tr>
<tr>
<td>BT2a</td>
<td>0.01 ± 0.006 (n = 3)</td>
<td>4.06 ± 1.39 (n = 3)</td>
<td>177</td>
</tr>
<tr>
<td>BT3a</td>
<td>0.01 ± 0.003 (n = 3)</td>
<td>3.95 ± 0.61 (n = 3)</td>
<td>409</td>
</tr>
<tr>
<td>BT4a</td>
<td>0.006 ± 0.006 (n = 3)</td>
<td>1.14 ± 0.91 (n = 3)</td>
<td>180</td>
</tr>
<tr>
<td>RSV-B</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>18537</td>
<td>0.4 ± 0.2 (n = 20)</td>
<td>2.4 ± 1.2 (n = 5)</td>
<td>6</td>
</tr>
<tr>
<td>B-TX-60567b</td>
<td>0.24 ± 0.22 (n = 3)</td>
<td>2.8 ± 0.8 (n = 3)</td>
<td>11.5</td>
</tr>
<tr>
<td>B-TX-61406b</td>
<td>0.028 ± 0.026 (n = 3)</td>
<td>2.0 ± 1.1 (n = 3)</td>
<td>72.3</td>
</tr>
<tr>
<td>B-TX-79233b</td>
<td>0.003 ± 0.001 (n = 3)</td>
<td>1.7 ± 2.2 (n = 3)</td>
<td>647.5</td>
</tr>
</tbody>
</table>

*IC_{50}, 50% inhibitory concentration; VN_{so}, 50% virus neutralization titer; n, number of isolates.

* These viruses have been reported previously (30,31).

100% with ALX-0171 and 84.4% with palivizumab (P = 0.053), and for RSV-B isolates, this proportion was 93.1% with ALX-0171 and 86.2% with palivizumab (P = 0.67). These results are consistent with published data demonstrating that palivizumab neutralized 75/77 (97%) clinical isolates tested (32). Importantly, ALX-0171 demonstrated a broad RSV strain neutralization capacity that was at least as extensive as that of palivizumab when looking at 2 log_{10} reductions.

Our data were the most striking when the complete suppression of RSV replication was compared between ALX-0171 and palivizumab. When they were tested at equivalent concentrations, ALX-0171 completely blocked replication in 87% of the viruses tested, whereas palivizumab completely blocked replication in 87% of the viruses tested, which is likely the consequence of the higher potency of ALX-0171 (as shown in Table 2) and the lower molecular weight of ALX-0171 compared with that of full-length MAbs (42.2 for ALX-0171 versus 148 for palivizumab [32]), is indicative of a lower neutralization threshold for ALX-0171 than for palivizumab.

**ALX-0171 binds to antigenic site II of F protein.** To identify the residues that are important for the binding of ALX-0171 to RSV F protein, studies of the binding of monovalent Nb017 to RSV-A Long escape mutants containing a single point mutation in antigenic site II or IV were performed (Fig. 2). These tested RSV escape mutants, which are listed in Table 4, were previously reported and selected with monoclonal antibodies to these sites (11, 28, 29). The binding of Nb017 to antigenic site II, but not site IV, mutants was significantly reduced (Table 4). In addition, binding of ALX-0171 to the RSV F protein competed with that of palivizumab, although only partially (Fig. 3 and data not shown). These results provide further evidence that Nb017 and, consequently, ALX-0171 target antigenic site II of the F protein and that its epitope is close to or overlaps that of palivizumab.

**ALX-0171 retains physicochemical properties following nebulization.** As the intended route of ALX-0171 administration is by inhalation, it is important that ALX-0171 withstand the nebulization process. An Akita2 Apixneb nebulizer was used to nebulize ALX-0171, and the collected aerosol was characterized for product stability by size-exclusion (SE) high-pressure liquid chromatography (HPLC) (to determine purity/impurity), reverse-phase (RP)-HPLC (to determine purity), and ELISA (to determine potency). These analytical methods were suitable for assessing higher-molecular-weight species in panels of RSV clinical isolates by palivizumab and ALX-0171 by 100-fold or greater and neutralization threshold for ALX-0171 than for palivizumab.

Table 3: Comparative inhibition of in vitro replication of a panel of RSV clinical isolates by palivizumab and ALX-0171 by ≥100-fold or completely

<table>
<thead>
<tr>
<th>RSV group</th>
<th>GMT^c (log_{10})</th>
<th>No. of isolates with ≥100-fold reduction/total no. of isolates tested (%)</th>
<th>P value^{d}</th>
<th>No. of isolates with complete inhibition/total no. of isolates tested (%)</th>
<th>P value^{e}</th>
</tr>
</thead>
<tbody>
<tr>
<td>RSV-A</td>
<td>4.9 ± 0.4</td>
<td>27/32 (84.4)</td>
<td>0.053</td>
<td>0/32 (0)</td>
<td>0.0001</td>
</tr>
<tr>
<td>RSV-B</td>
<td>4.7 ± 0.4</td>
<td>25/29 (86.2)</td>
<td>0.67</td>
<td>11/29 (37.9)</td>
<td>0.003</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td>52/61 (85.2)</td>
<td>0.054</td>
<td>11/61 (18)</td>
<td>0.0001</td>
</tr>
</tbody>
</table>

^c GMT, geometric mean titer of buffer controls.

^d Differences between groups were analyzed using Fisher's exact test for comparison of proportions. Significance was defined at a P value of <0.05.

^e Complete virus inhibition was defined as no detectable virus plaques. The limit of virus detection in the plaque reduction neutralization assay was 5 PFU/ml.
was the difference in F-protein expression. Nb017 was used at 0.2 antigenic site I (191C7) with preserved binding to all depicted mutants to account for.

The residues listed are those that were mutated in the tested RSV escape mutants and are shown in yellow. The figure was prepared by using ICM Molsoft (46) and was derived from the sequence with PDB accession number 4JHW (26).

Furthermore, no effect of nebulization on ALX-0171 potency was observed by ELISA analysis (see Table S2 in the supplemental material).

Local delivery of ALX-0171 to cotton rats reduces the RSV load in the nose and lung. Cotton rats were used to evaluate the in vivo efficacy of ALX-0171 against RSV. Different doses of ALX-0171 were administered via the intranasal route on different days after RSV Tracy challenge either once (day 2 or day 3) or twice (day 2 and day 3) or by nebulization 1 h before RSV Tracy challenge (Table 5). On day 4 the cotton rats were sacrificed and the viral titers in both the nose and lung were determined.

Intranasal administration of ALX-0171 on day 2, day 3, or days 2 and 3 after RSV challenge resulted in significant viral load reductions in the lungs ranging from ~50-fold (1.66 log<sub>10</sub>) for the lowest dose tested (1 mg/kg) (P < 0.0001) to ~2,000-fold (3.3 log<sub>10</sub>) for the highest dose tested (68 mg/kg) (P < 0.0001). Nasal RSV titers were also significantly reduced for all doses and schedules tested (P < 0.0001), except for the day 2 treatment at 4 mg/kg (P = 0.7).

As the intended route of delivery of ALX-0171 is by nebulization, we performed a proof-of-concept experiment to confirm that nebulization did not affect the antiviral activity of ALX-0171. For this experiment, prophylactic delivery of ALX-0171 via nebulization was chosen as the most straightforward approach. In total, 4 groups of 6 cotton rats received estimated delivered doses of 0.3 mg/kg, 0.8 mg/kg, and 2 mg/kg 1 h before RSV Tracy challenge (Table 5). A dose-dependent reduction in viral titers was seen in the nose, which was significant for the two higher doses (P = 0.01 and 0.006 for doses of 0.8 mg/kg and 2 mg/kg, respectively). Furthermore, RSV replication was almost completely blocked in the lungs, even at the lowest dose tested (>3.67 log<sub>10</sub> reduction, P < 0.0001).

DISCUSSION

ALX-0171 is a novel therapeutic biologic in development for the treatment of RSV infections in infants. ALX-0171 is a trimeric Nanobody that binds an epitope in antigenic site II of RSV F protein with subnanomolar affinity. This epitope partially overlaps the palivizumab epitope.

The formatting of the monovalent Nb017 into the trivalent Nanobody ALX-0171 increased the potency of neutralization (>6,000-fold) of both RSV-A and RSV-B strains. Although formatting clearly increased the binding affinity for the biologically relevant prefusion conformation of the F protein (~160-fold), it does not fully explain the observed large increase in the virus neutralization capacity of ALX-0171. Our results are consistent with those of a previous report demonstrating that a bivalent Nanobody construct specific for F-protein antigenic site II had a 4,000-fold improved neutralization capacity compared to that of the monovalent construct against RSV-A Long (22). Likewise, the difference in potency between palivizumab Fab and the full-length antibody is 100-fold, while there is only a minimal improvement
in the $K_d$ (33). However, the basis of this observation is not yet understood, and the precise mechanisms involved remain under investigation.

Like palivizumab, Nanobodies Nb017 and ALX-0171 bind to both the postfusion and prefusion conformation of the F protein and, as a result, likely inhibit the conformational changes related to F-protein activation, as suggested for other Nanobodies and monoclonal antibodies specific for antigenic site II (21, 34–36). This hypothesis is supported by a model of the RSV F-protein prefusion structure, in which the residues of antigenic site II are extended complementarity-determining region loops may enter 

**TABLE 5 Reduction in RSV replication in cotton rats**

<table>
<thead>
<tr>
<th>Expt no., route of delivery</th>
<th>ALX-0171 dose delivered (mg/kg)</th>
<th>Treatment regimen</th>
<th>Nasal titer Mean log$<em>{10}$ total no. PFU (± SD) vs buffer Log$</em>{10}$ reduction vs buffer</th>
<th>Lung titer Mean log$<em>{10}$ no. of PFU/g lung (± SD) vs buffer Log$</em>{10}$ reduction vs buffer</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Expt 1, intranasal</strong></td>
<td>Buffer</td>
<td>Days +2 and +3</td>
<td>4.78 (0.40)</td>
<td>4.94 (0.13)</td>
</tr>
<tr>
<td></td>
<td>1</td>
<td>Days +2 and +3</td>
<td>3.46 (0.56)</td>
<td>1.32*</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>Days +2 and +3</td>
<td>3.06 (0.75)</td>
<td>1.72*</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>Days +2 and +3</td>
<td>2.53 (0.87)</td>
<td>2.26*</td>
</tr>
<tr>
<td></td>
<td>21</td>
<td>Days +2 and +3</td>
<td>1.68 (0.28)</td>
<td>3.10*</td>
</tr>
<tr>
<td><strong>Expt 2, intranasal</strong></td>
<td>Buffer</td>
<td>Days +2 and +3</td>
<td>4.84 (0.15)</td>
<td>5.19 (0.24)</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>Days +2 and +3</td>
<td>2.00 (0.72)</td>
<td>2.84*</td>
</tr>
<tr>
<td></td>
<td>19</td>
<td>Days +2 and +3</td>
<td>1.73 (0.21)</td>
<td>3.31*</td>
</tr>
<tr>
<td></td>
<td>68</td>
<td>Days +2 and +3</td>
<td>1.54 (0.59)</td>
<td>3.30*</td>
</tr>
<tr>
<td><strong>Expt 3, intranasal</strong></td>
<td>Buffer</td>
<td>Days +2 and +3</td>
<td>4.81 (0.36)</td>
<td>4.88 (0.34)</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>Day +2</td>
<td>4.53 (0.87)</td>
<td>0.28</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>Day +3</td>
<td>1.84 (0.38)</td>
<td>2.97*</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>Days +2 and +3</td>
<td>1.74 (0.30)</td>
<td>3.07*</td>
</tr>
<tr>
<td><strong>Expt 4, nebulization</strong></td>
<td>Buffer$^a$</td>
<td>Hour −1</td>
<td>5.04 (0.25)</td>
<td>5.05 (0.22)</td>
</tr>
<tr>
<td></td>
<td>0.3$^b$</td>
<td>Hour −1</td>
<td>3.97 (0.62)</td>
<td>1.07</td>
</tr>
<tr>
<td></td>
<td>0.8$^b$</td>
<td>Hour −1</td>
<td>3.02 (1.46)</td>
<td>2.02*</td>
</tr>
<tr>
<td></td>
<td>2$^b$</td>
<td>Hour −1</td>
<td>2.87 (1.42)</td>
<td>2.18*</td>
</tr>
</tbody>
</table>

$^a$ Cotton rats (6 to 8 rats per group) were challenged with 1.41 × 10$^3$ to 3.4 × 10$^3$ PFU of RSV Tracy (intranasally, 100 μl) on day 0. The animals received ALX-0171 at the indicated times either by intranasal administration or by nebulization. Viral titers and viral RNA loads in both the lung and nose were assessed on day 4 postinfection. *, P < 0.05 versus buffer control. P values were determined by ANOVA with a follow-up Dunnett’s pairwise comparison test.

$^b$ The delivered dose was estimated. The estimated delivered dose was defined as the estimated dose that is inhalable by the cotton rats and was calculated using the following formula: delivered dose (in micrograms per kilogram of body weight) = (aerosol concentration [in micrograms per liter]) × (minute volume [in liters-minute per kilogram]) × (treatment time [in minutes]) × percentage of particles of ≤5 μm (respirable fraction), where the minute volume was estimated to be 0.7 liter-min/kg (47) and the respirable fraction was estimated to be 80% on the basis of the characteristics of the nebulizer.

independent F-protein trimers or simultaneous binding of the three subunits within a single F-protein trimer (20, 21), as the length of the GS linkers was specifically designed to allow such interactions. Further crystallography studies are needed to elucidate the mechanisms involved in this increased neutralization capacity of ALX-0171.

**In vivo** ALX-0171 was shown to be highly efficient at reducing and/or blocking RSV replication in the lung and nose, similar to what was reported for motavizumab, an affinity-matured version of palivizumab (39, 40), whereas no effect on viral load in the nose was demonstrated for palivizumab at doses of ≤15 mg/kg (40, 41). There remains the possibility that the residual ALX-0171 present in the wash fluid interfered in the plaque assay. Nonetheless, despite this caveat, we believe that it remains the most valid assessment, as it reflects the fact either that less virus is present or that any virus still present is effectively neutralized. Importantly, as ALX-0171 is devoid of an Fc, its in vivo efficacy likely relies entirely on direct antiviral activity, in contrast to what has been reported for palivizumab (42). In addition and in contrast to the route of delivery of palivizumab and motavizumab, which were administered systemically in therapeutic trials in infants and demonstrated conflicting effects on nasal/tracheal viral loads (39, 41, 43), ALX-0171 is delivered straight to the site of infection. Direct administration to the airways is likely to provide faster and more robust antiviral activity in the respiratory tract, which may be critical for the treatment of an acute disease like RSV infection. Indeed, the therapeutic effect of topical administration of purified human immunoglobulins, screened for high RSV neutralization
activity, resulting in a decrease in viral loads in cotton rats was shown to be 160 times greater than that by administration by the parenteral route (44). In addition to the greater effectiveness of the ALX-0171 administration route, the neutralization threshold of ALX-0171 (i.e., the expected lung concentration needed to exert the full antiviral effect) is also anticipated to be lower than that of palivizumab. This is particularly important in a clinical setting where the administered dose is a limiting factor.

In summary, ALX-0171 represents a novel, highly potent anti-viral with broad specificity toward a large panel of RSV clinical isolates and may have significant potential for therapeutic use. Furthermore, direct delivery of ALX-0171 to the airways/lungs by nebulization proved an effective mode of drug delivery, as even the lowest dose of only 1 mg/kg delivered intranasally still showed antiviral efficacy. As nebulization has been shown to result in fast and efficient drug delivery to the principal site of RSV infection (45), i.e., the upper and lower respiratory tract, this mode of drug delivery may provide major therapeutic advantages for ALX-0171 in treating RSV-infected patients.

ACKNOWLEDGMENTS

We thank Valerie Lambert, Maureen Van den Hemel, Jorn Audiens, and Ananza Vandervorst for their technical support and expertise, Veronique De Brabandere and her team for the Nanobody physicochemical characterization, Erika Morizzo for providing the structural model of the prefusion F protein, and Hans Ulrichs and his team for fruitful scientific discussions. We also thank Vectura GmbH for the supply of the nebulizers. L.D., C.S., K.A., and E.D. are all current employees of Ablynx; all employees own stock/stock options of Ablynx. T.S. was an employee of Ablynx at the time of data generation. C.P., P.A.P., B.E.G., V.M., A.M., U.F.P. and J.A.M. undertook this work as part of a research contract with Ablynx.

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FUNDING INFORMATION

This work was supported by the Agentschap voor Innovatie door Wetenschap en Techniek (IWT), Belgium (grant numbers 100353 and 130562). Work in Madrid was partially supported by grant SAF2012-31217 to J.A.M. from Plan Nacional I+D+i.

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23. National Institutes of Health. 18 June 2015, posting date. A multicentre study in otherwise healthy infants and toddlers hospitalized for and diagnosed with RSV lower respiratory tract infection to evaluate the safety, tolerability, and clinical activity of ALX-0171. National Institutes of Health, Bethesda, MD.


